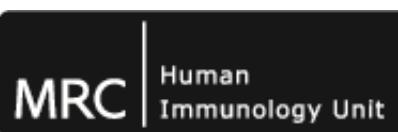




Super-resolution STED microscopy of the living cell



Weatherall
Institute of
Molecular
Medicine

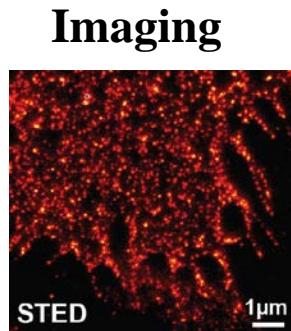
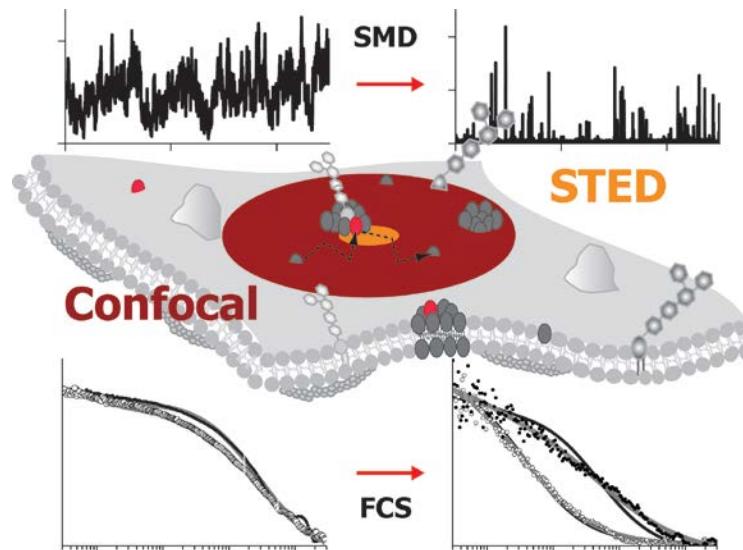
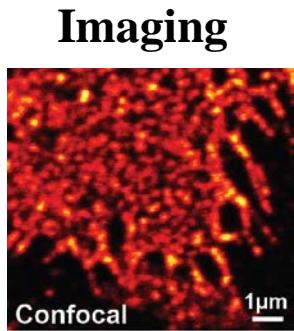


Christian Eggeling

Weatherall Institute of Molecular Medicine, HIU
University of Oxford

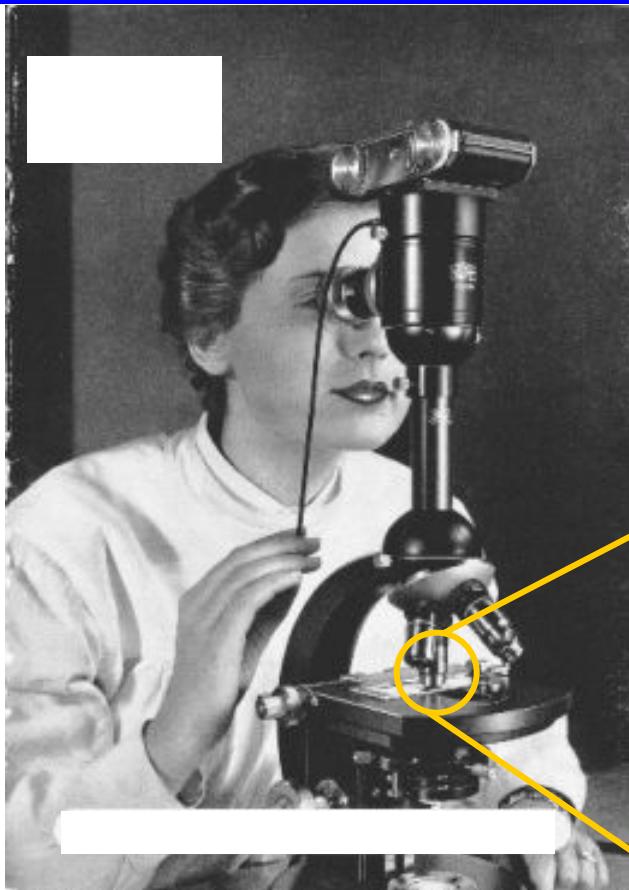
Previously:

Max Planck Institute for biophysical Chemistry
Dep. NanoBiophotonic (Prof. Hell)
Göttingen, Germany

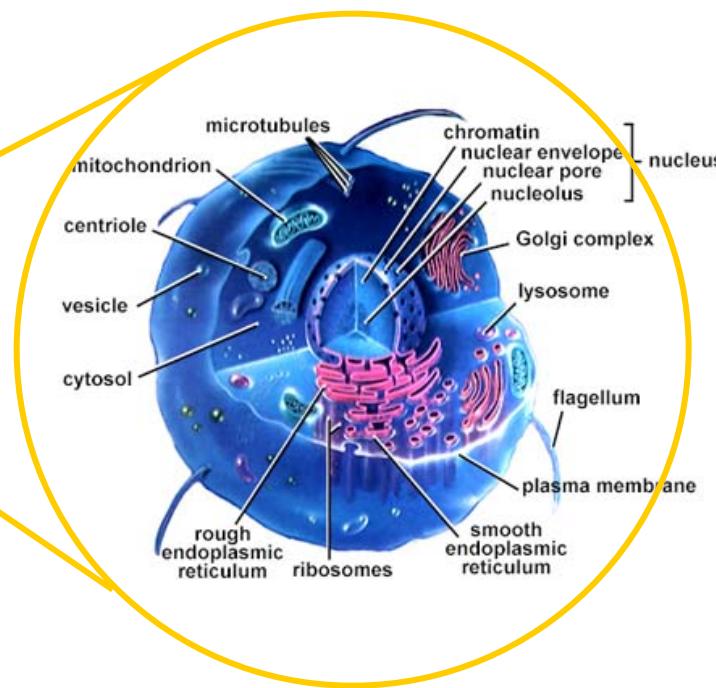
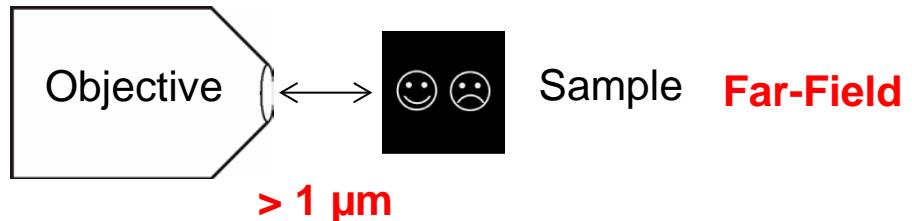


Live Cell Microscopy

Observation of living cells: Non-Invasive



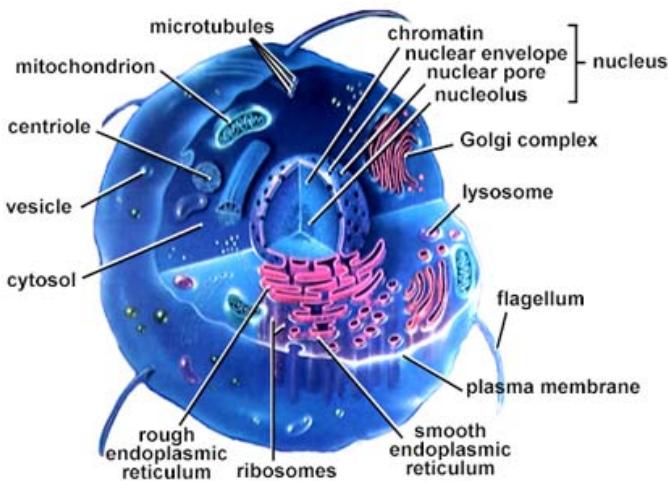
Light + Far-Field: non-invasive!



Live Cell Far-Field Microscopy

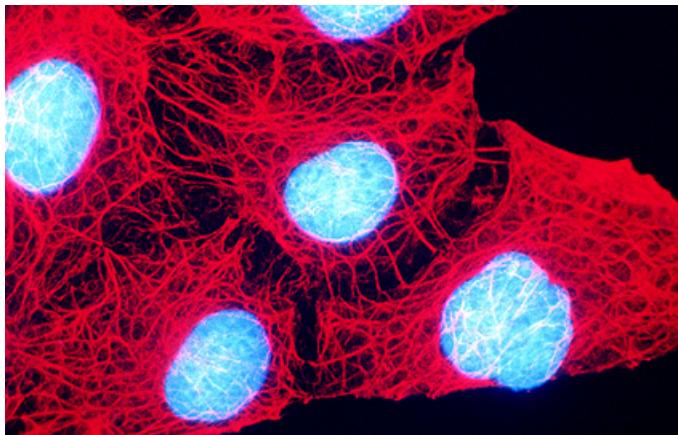
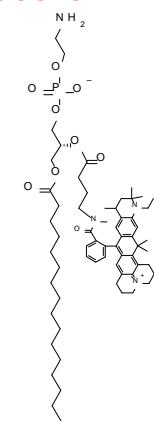
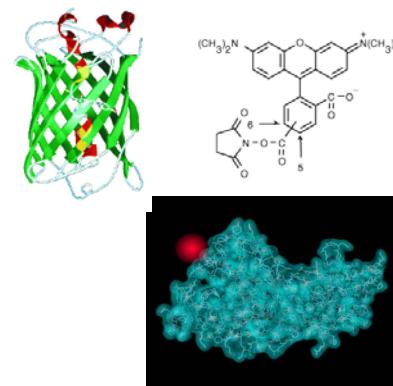
Fluorescence

Study specific molecular processes in the living cell:

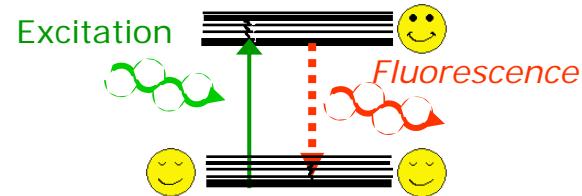


Fluorescence microscopy

Label specific protein/molecule



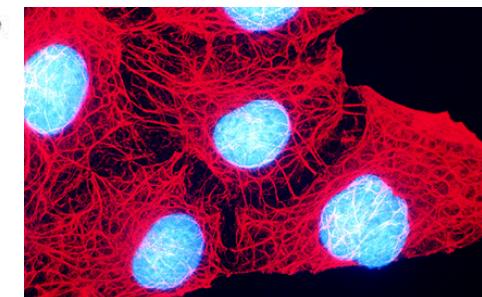
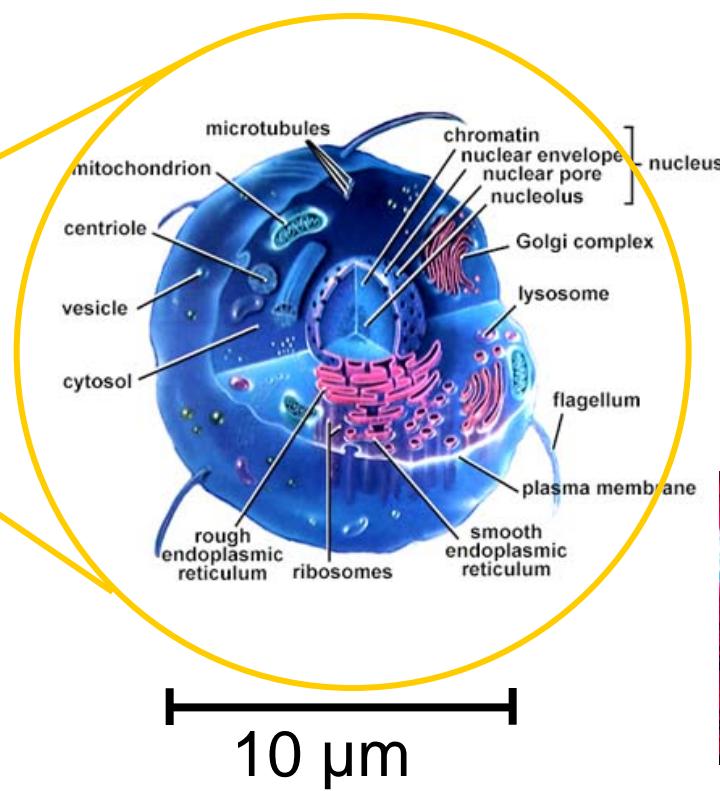
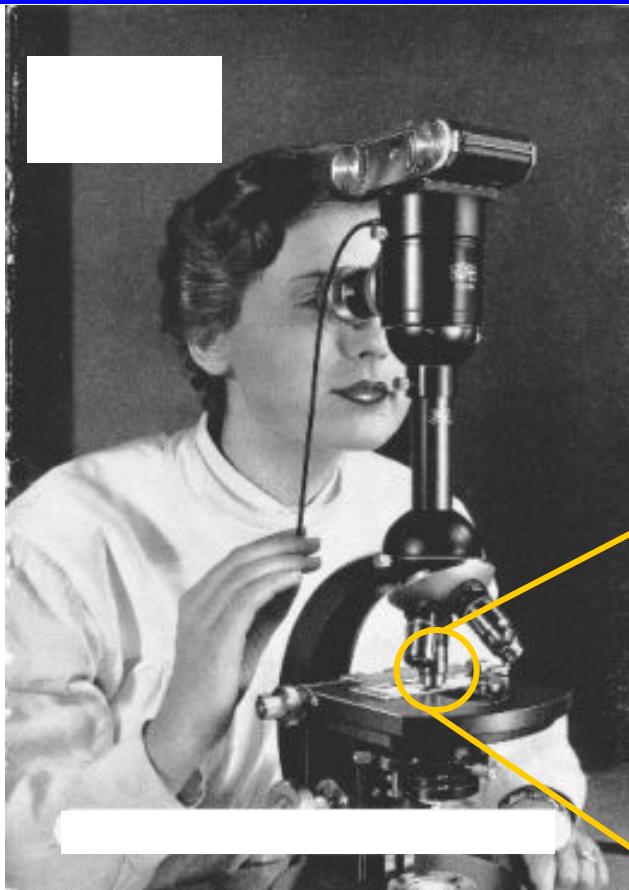
Excite fluorescence by laser light



Liver-Cells: Nucleus and Cell-skeleton

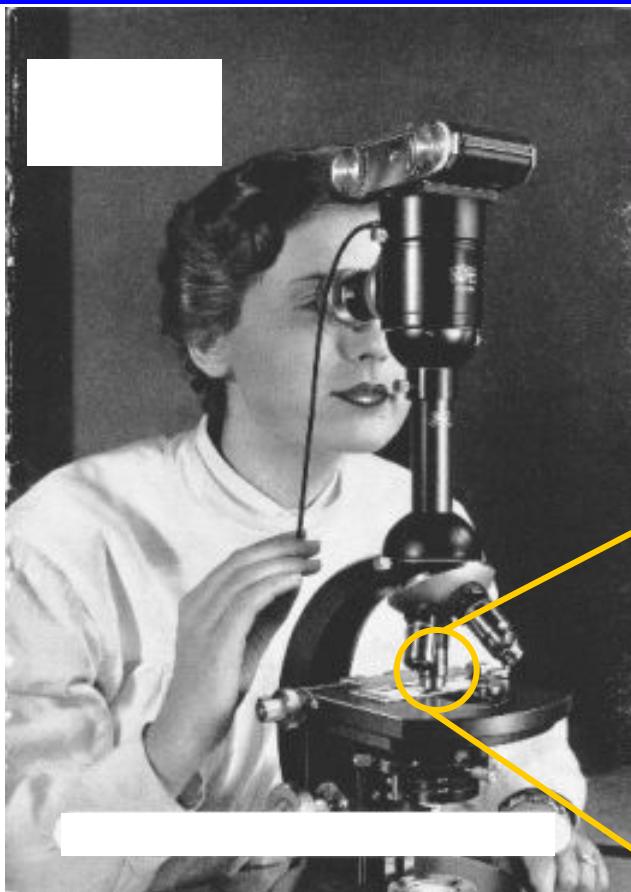
Far-Field Fluorescence Microscopy

Resolution: Goal

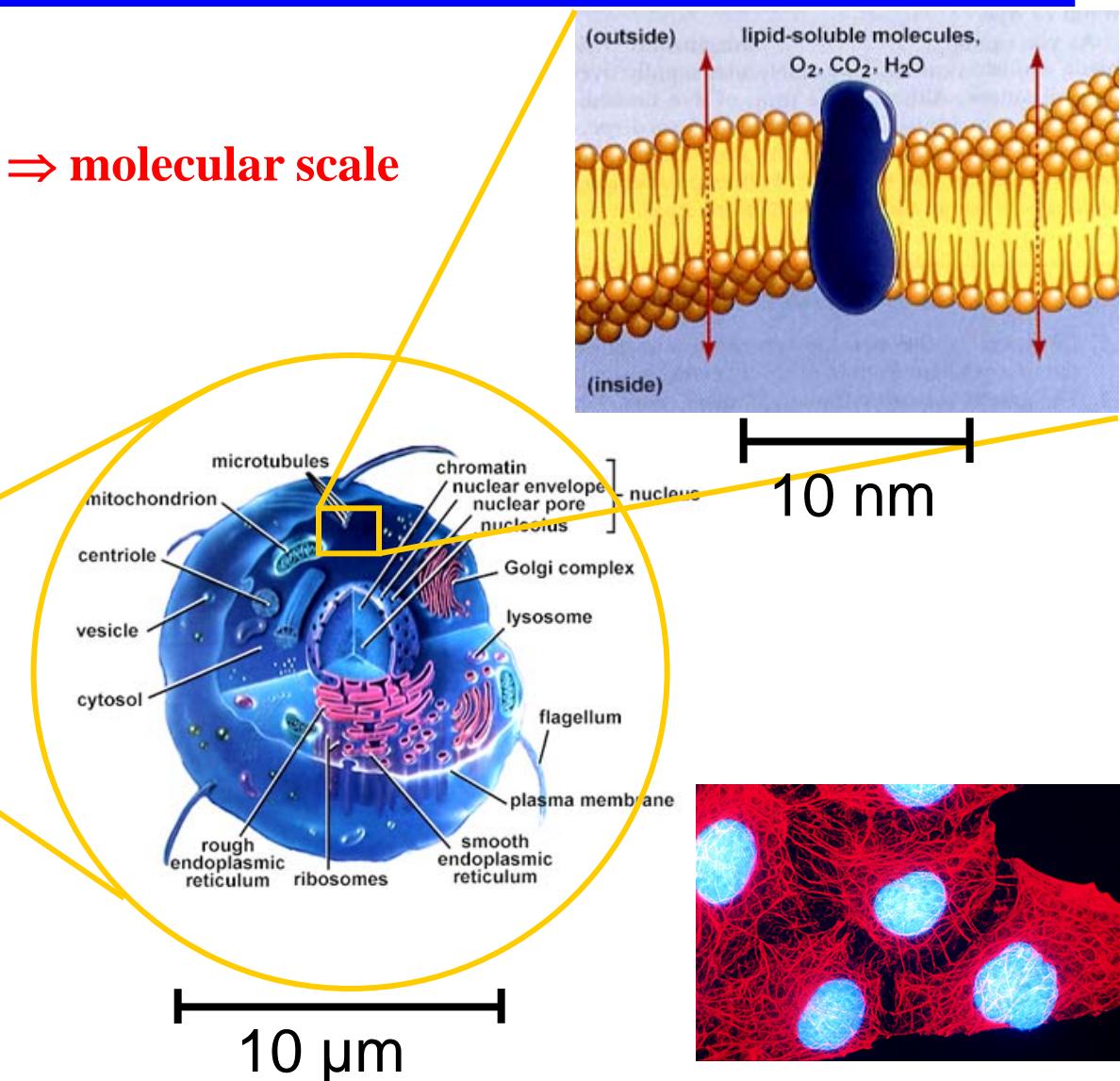


Far-Field Fluorescence Microscopy

Resolution: Goal



⇒ molecular scale

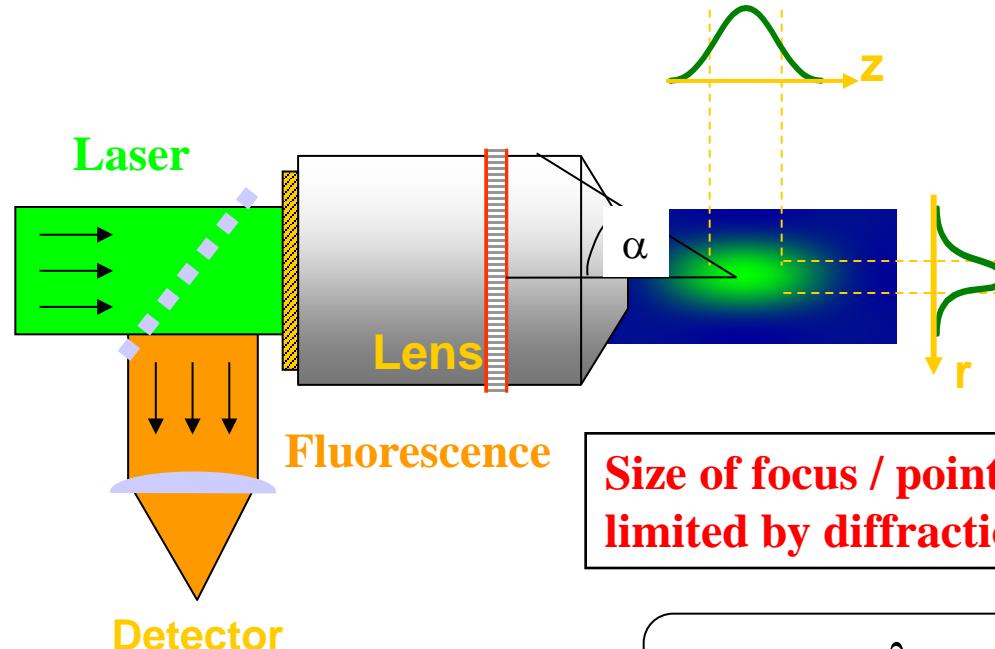


Far-Field Microscopy

Resolution Limit: Diffraction Barrier

Far-Field Fluorescence Microscopy: Focussing of light

- away from surfaces – inside cells (3D)



Size of focus / point-spread function limited by diffraction of light!!!

$$\Delta x = \frac{\lambda}{2n \sin \alpha}$$

Ernst Abbe 1873

Far-Field Microscopy

Resolution Limit: Diffraction Barrier

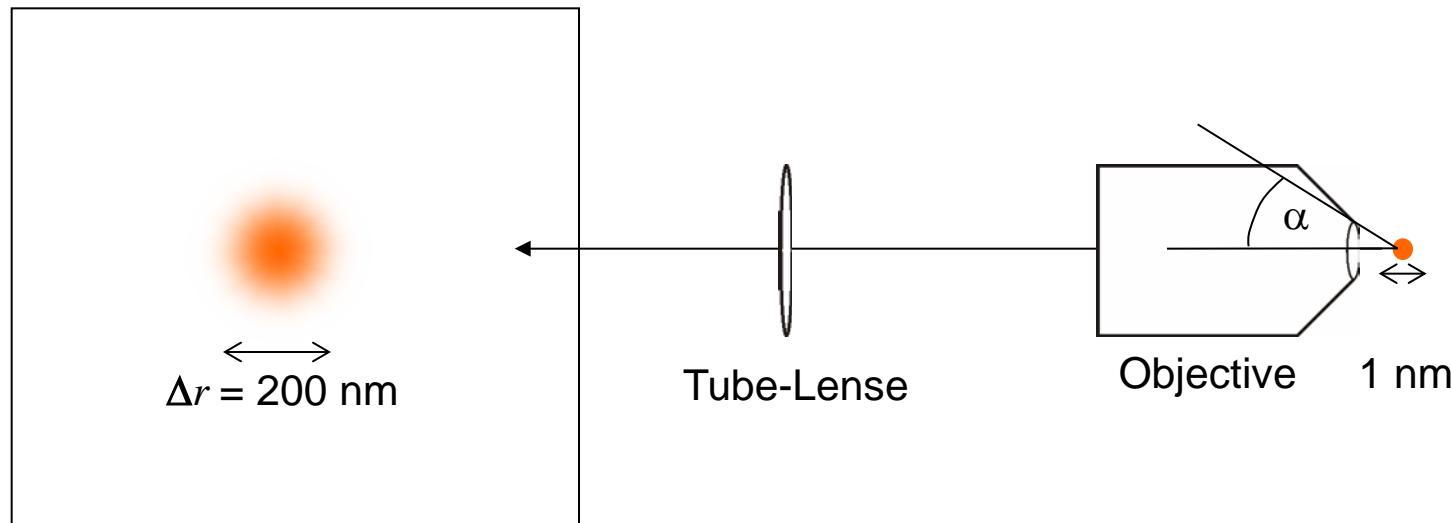
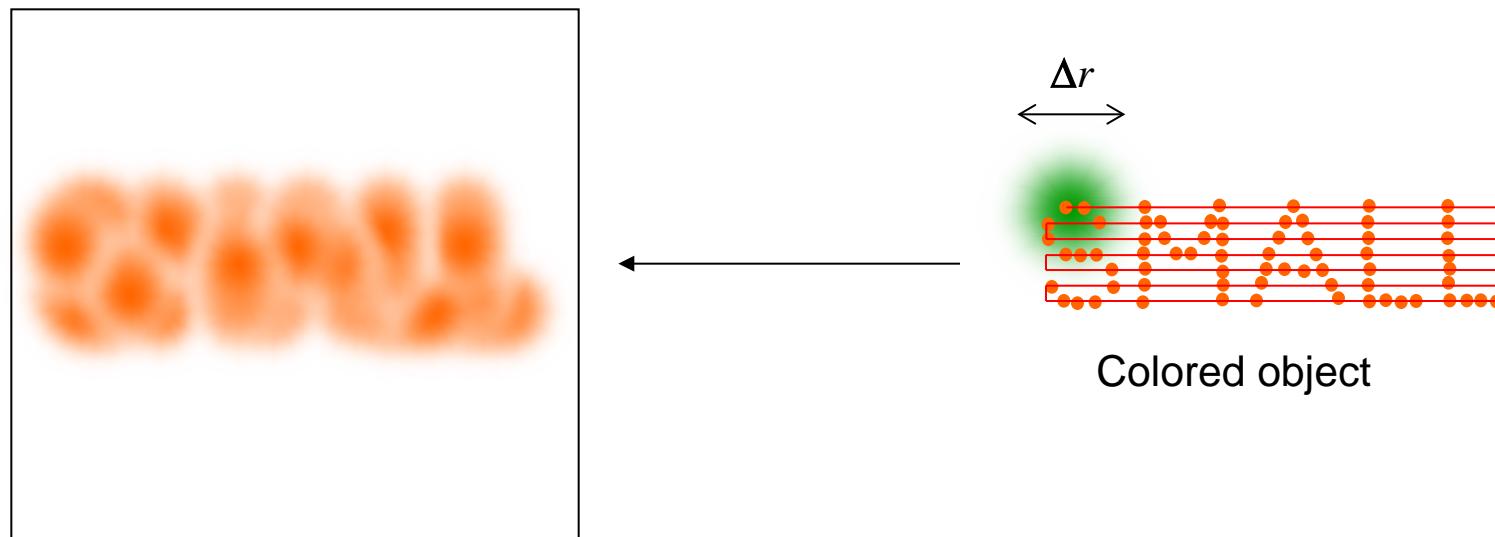
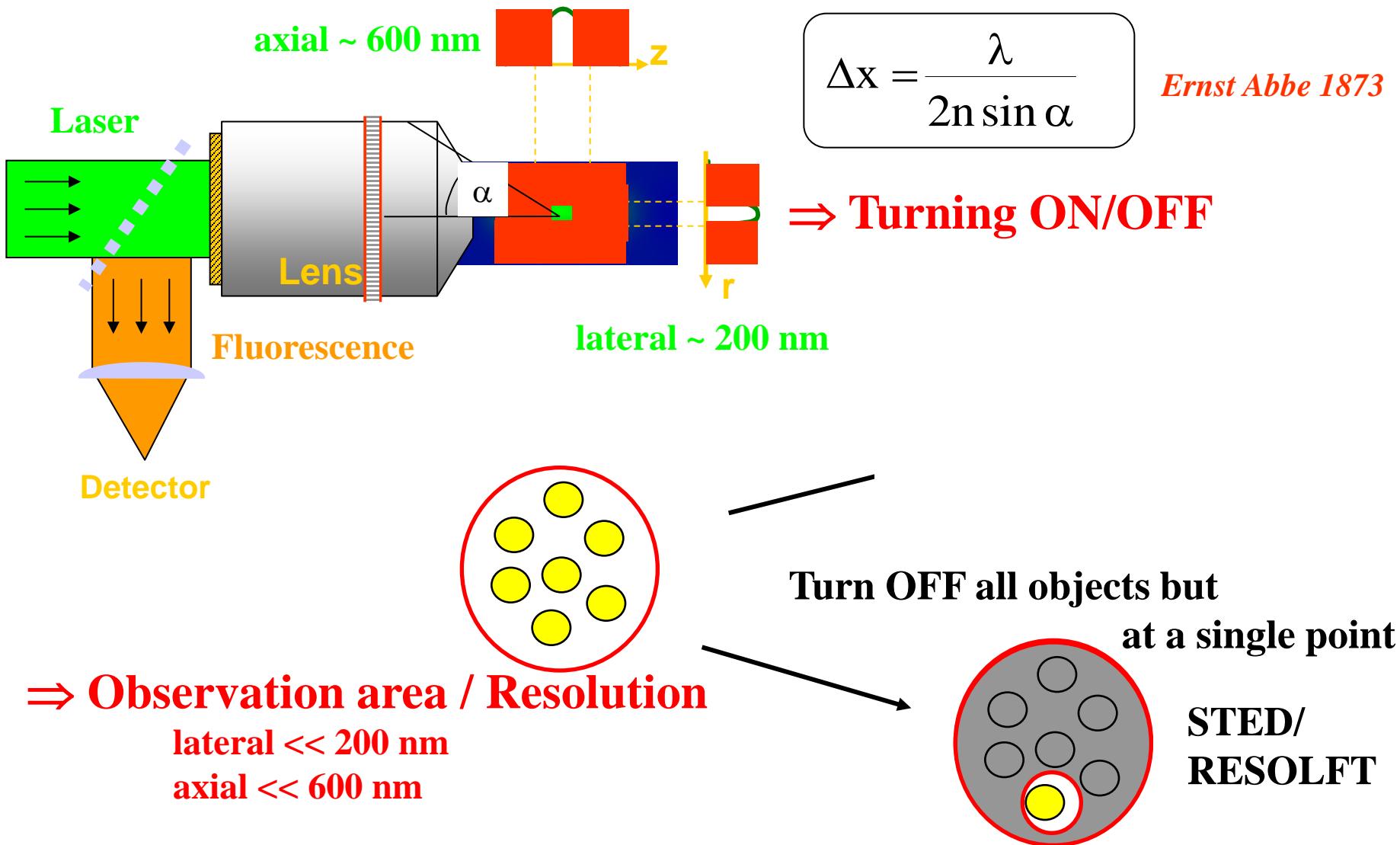


Image: diffraction-limited



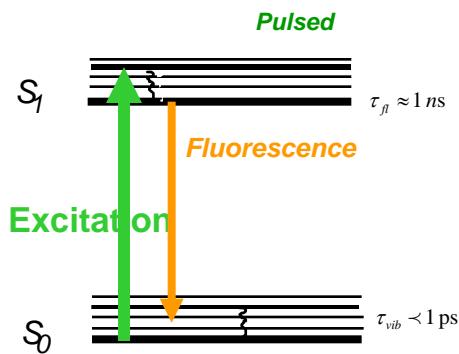
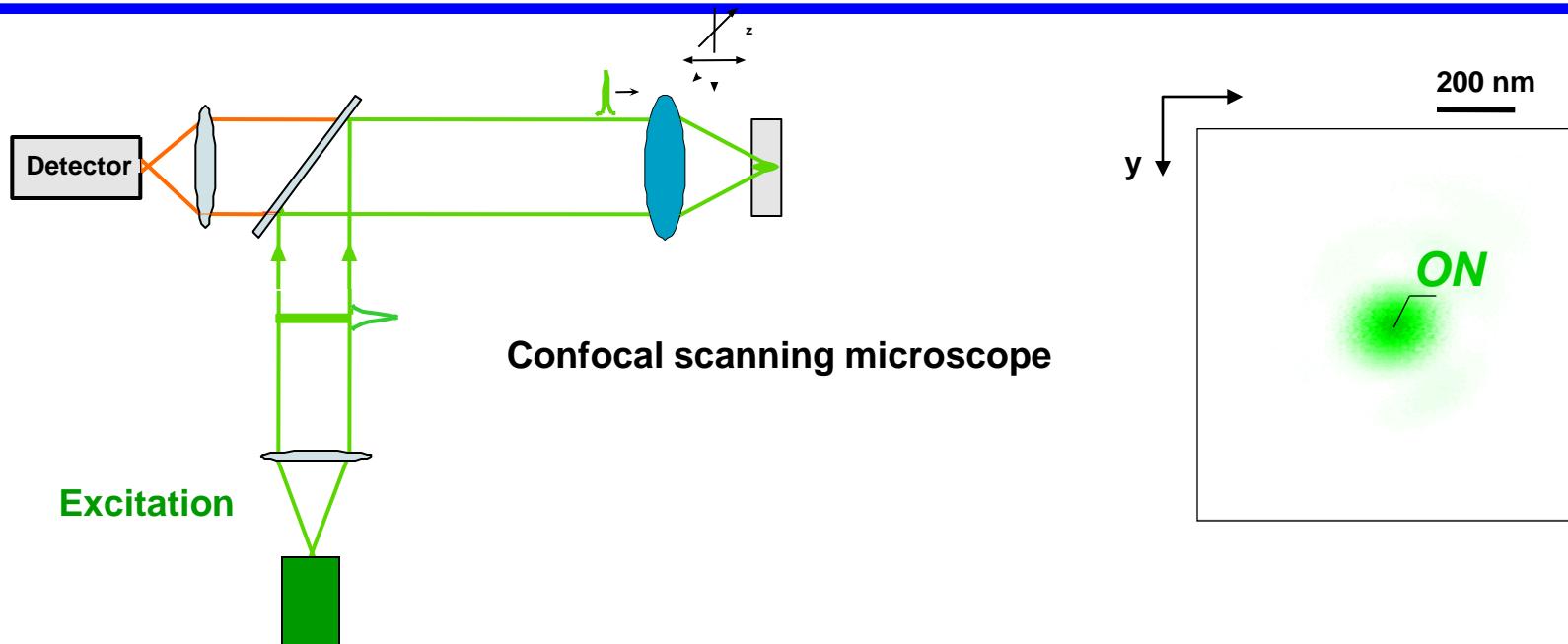
Far-Field Microscopy

Surpassing the Resolution Limit: Turning ON/OFF



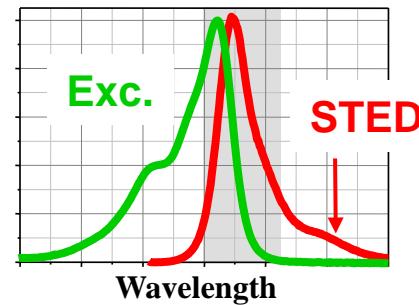
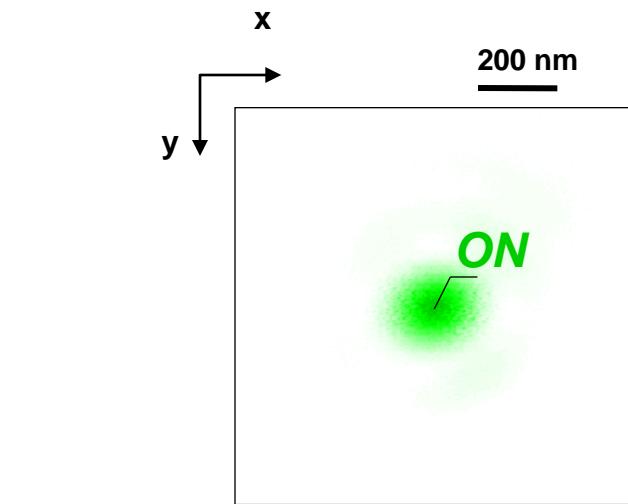
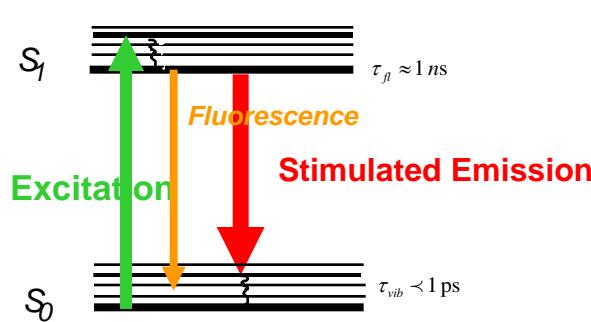
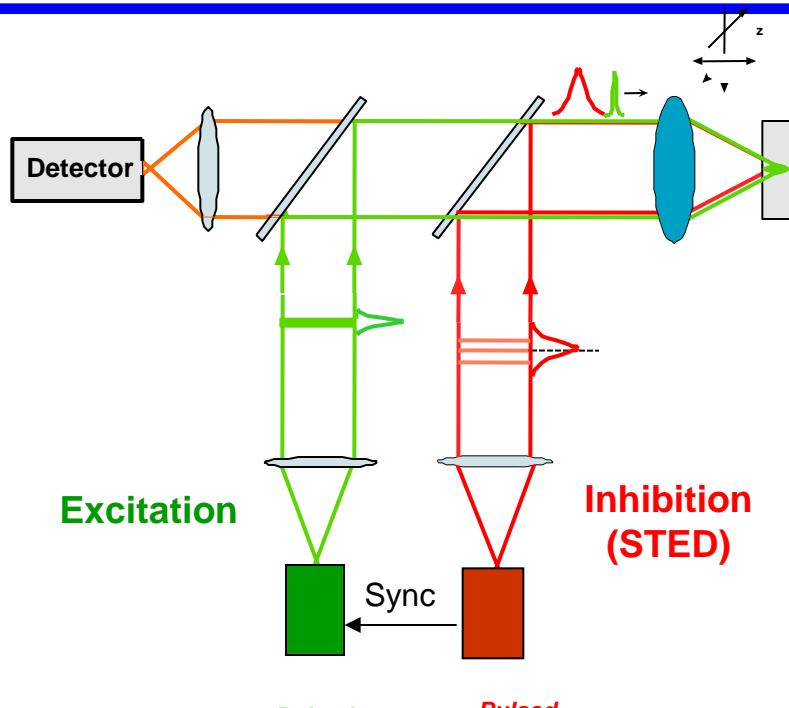
Fluorescence Microscopy

STED Microscopy



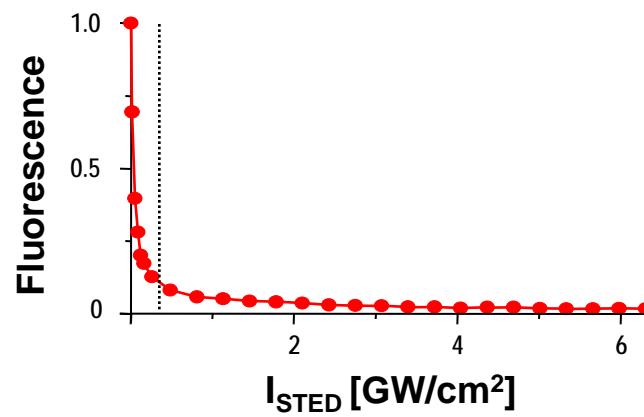
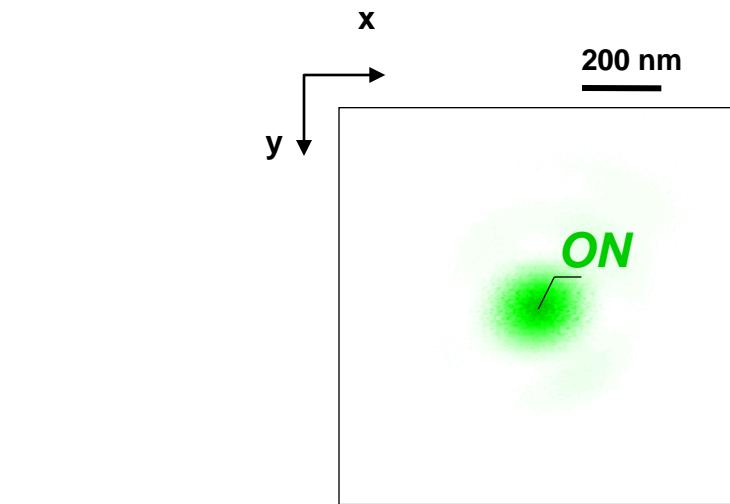
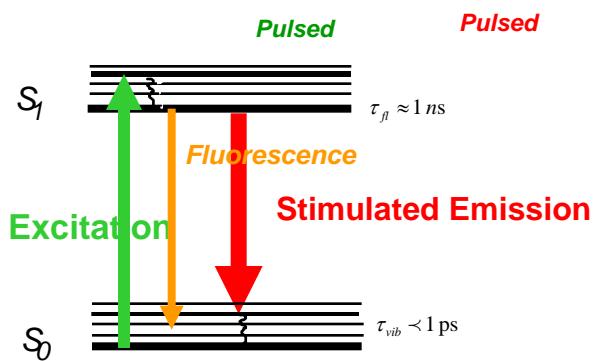
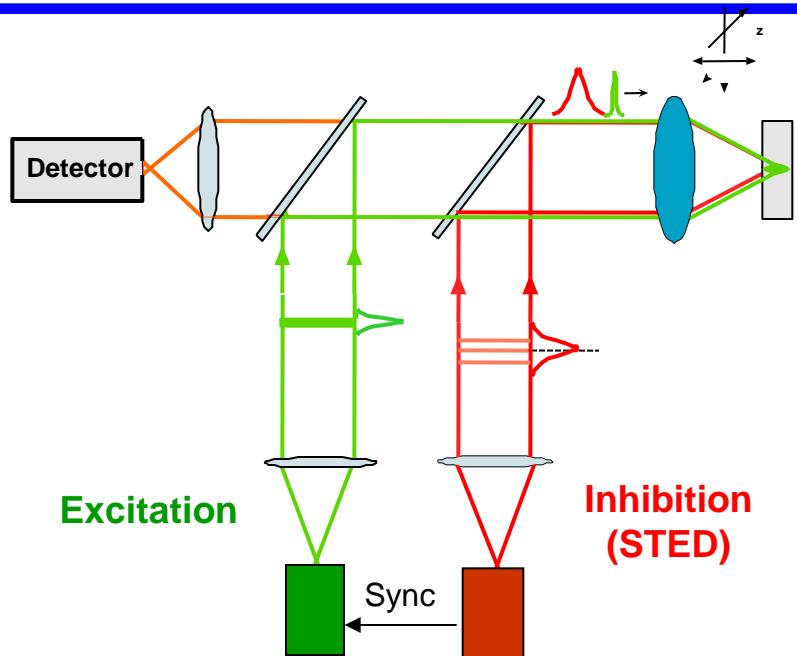
Fluorescence Microscopy

STED Microscopy



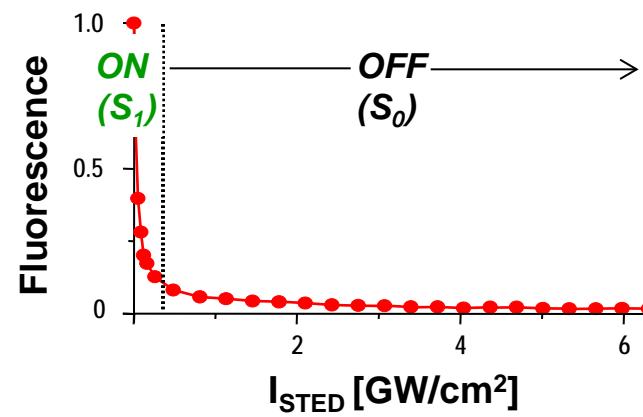
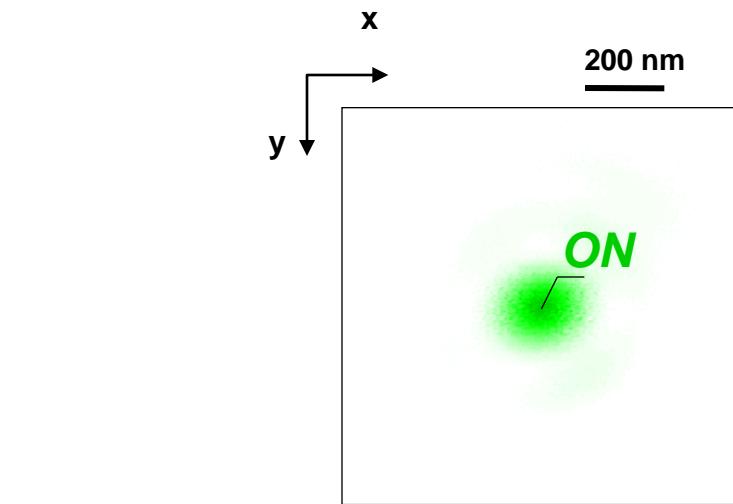
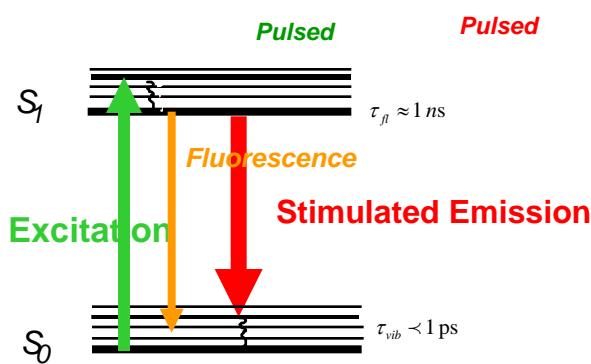
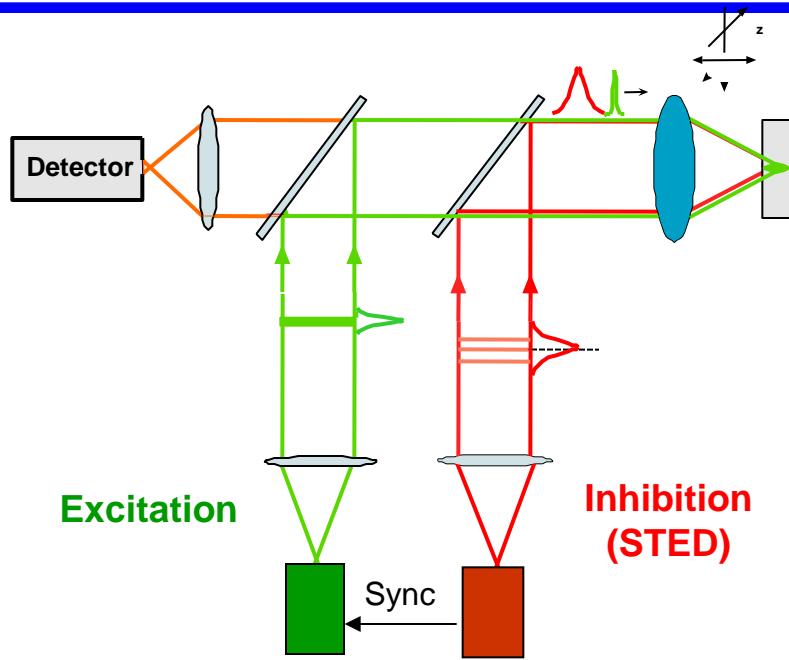
Fluorescence Microscopy

STED Microscopy



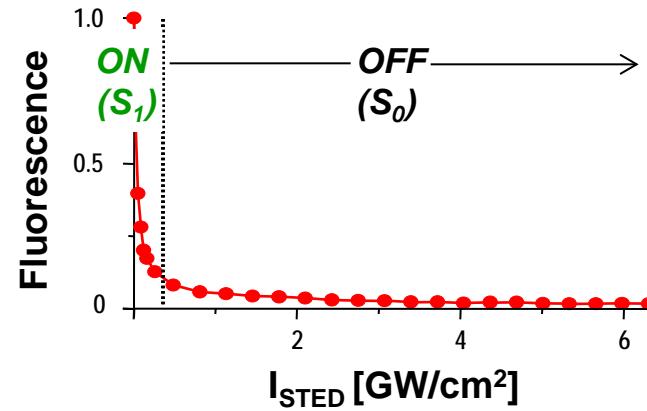
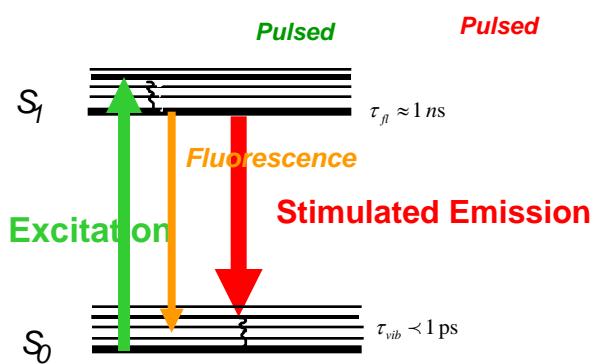
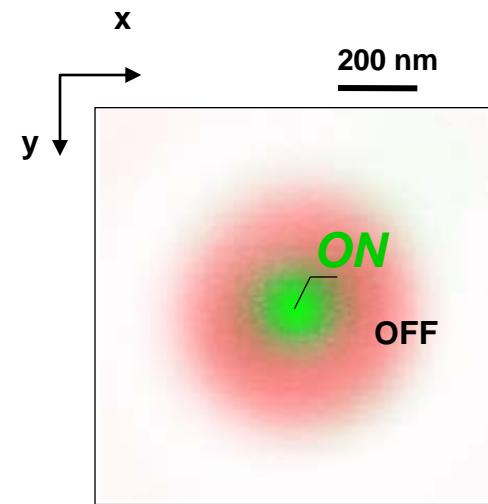
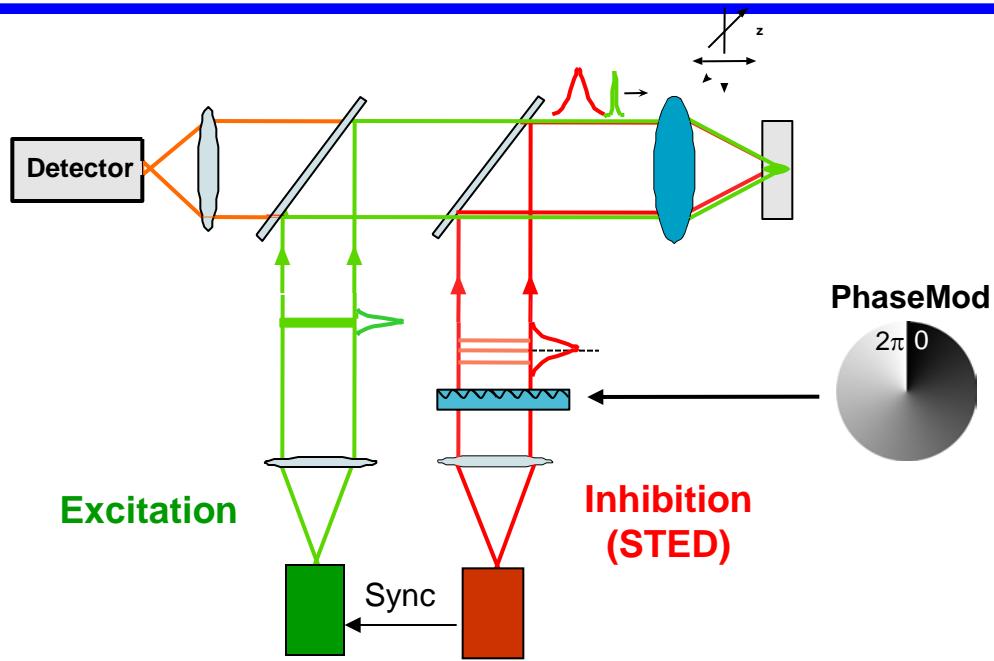
Fluorescence Microscopy

STED Microscopy



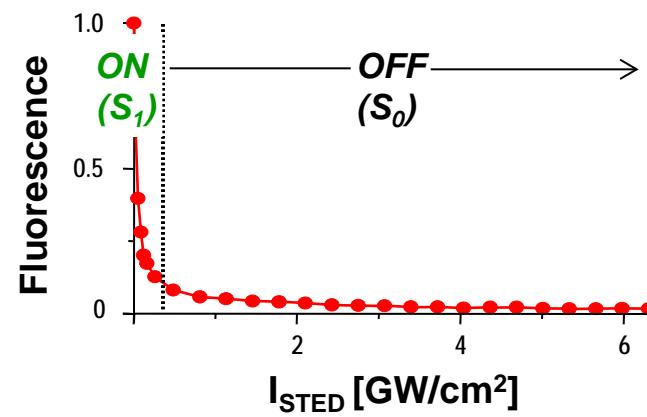
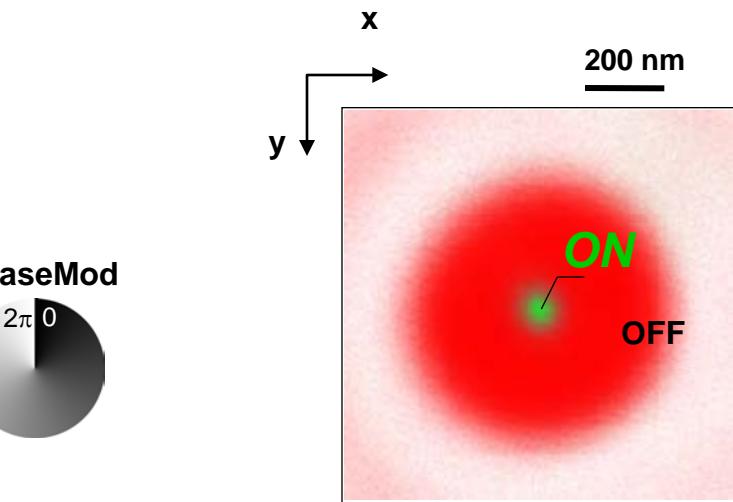
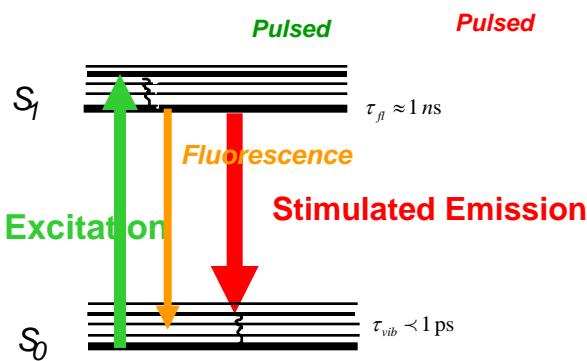
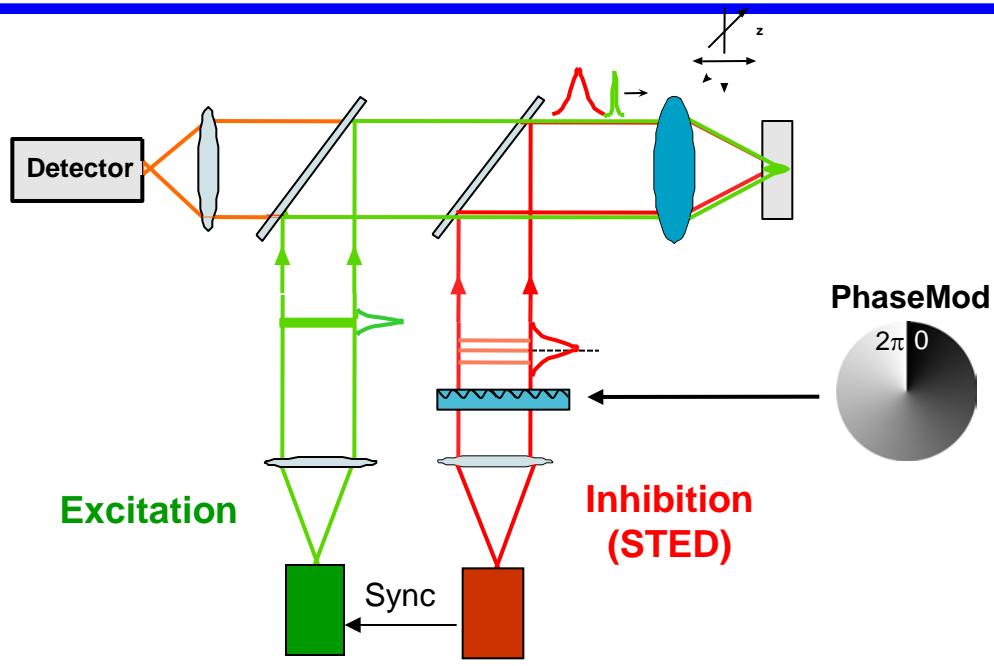
Fluorescence Microscopy

STED Microscopy



Fluorescence Microscopy

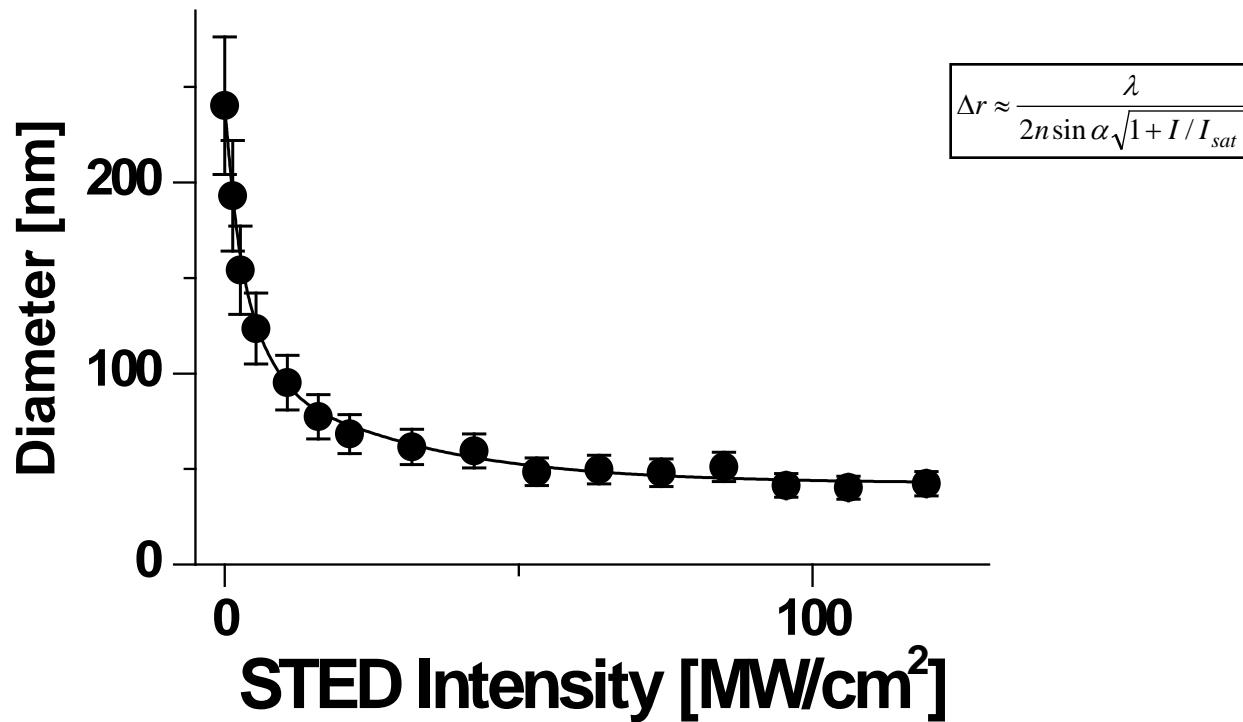
STED Microscopy



STED Microscopy

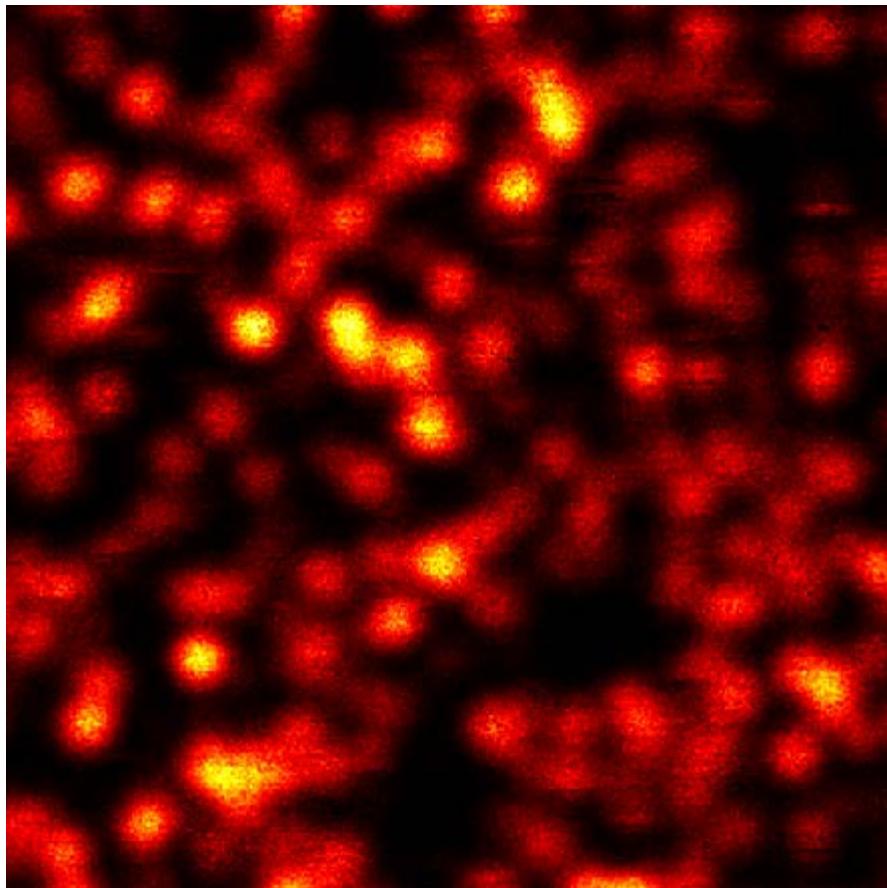
Dynamical confinement of resolution

Nanoscale observation areas: CONTINUOUS TUNING of spatial resolution!

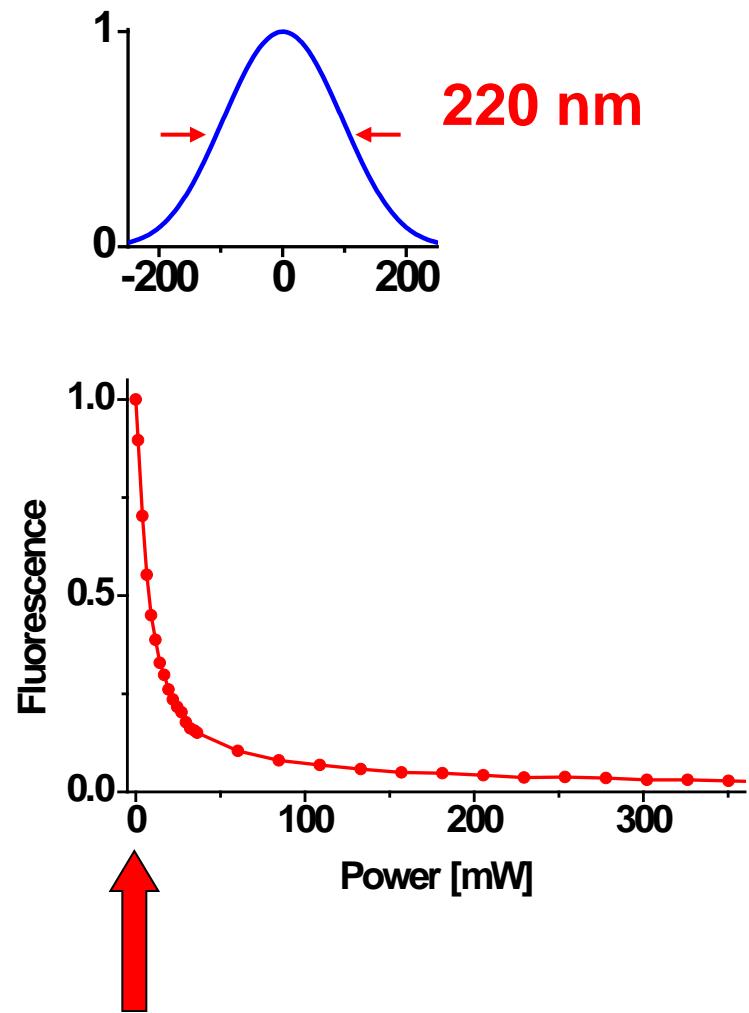


STED-Microscopy

Sub-Diffraction Imaging

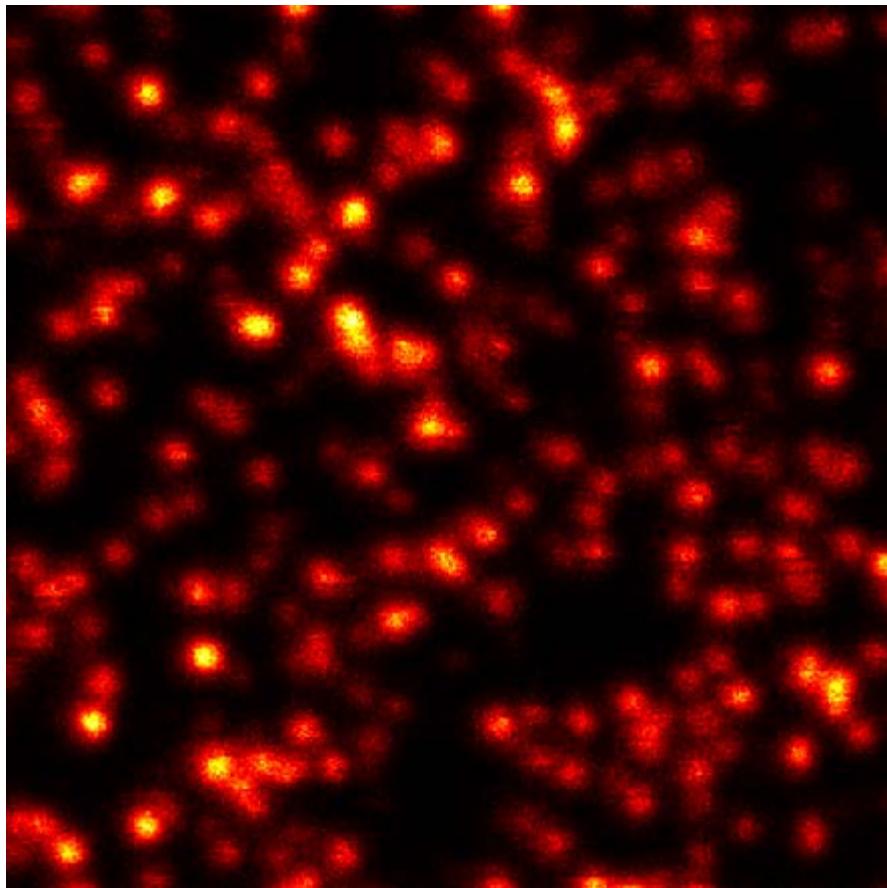


20nm Crimson beads
633nm exc, 90ps, 30kW/cm²
785nm STED 200ps, 76MHz

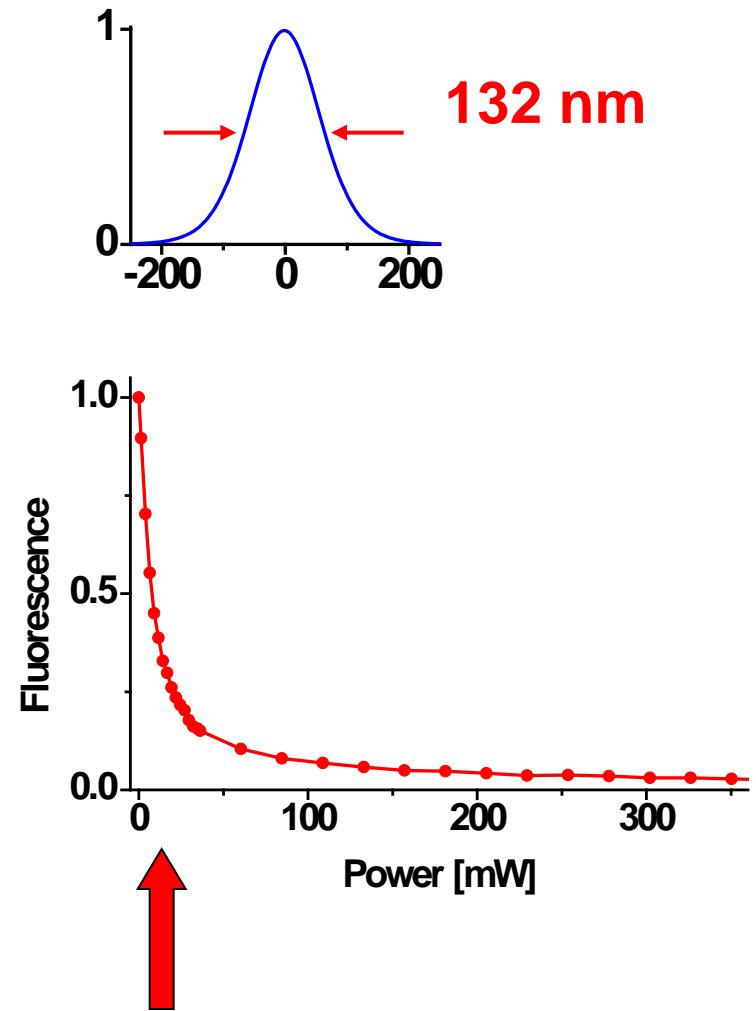


STED-Microscopy

Sub-Diffraction Imaging

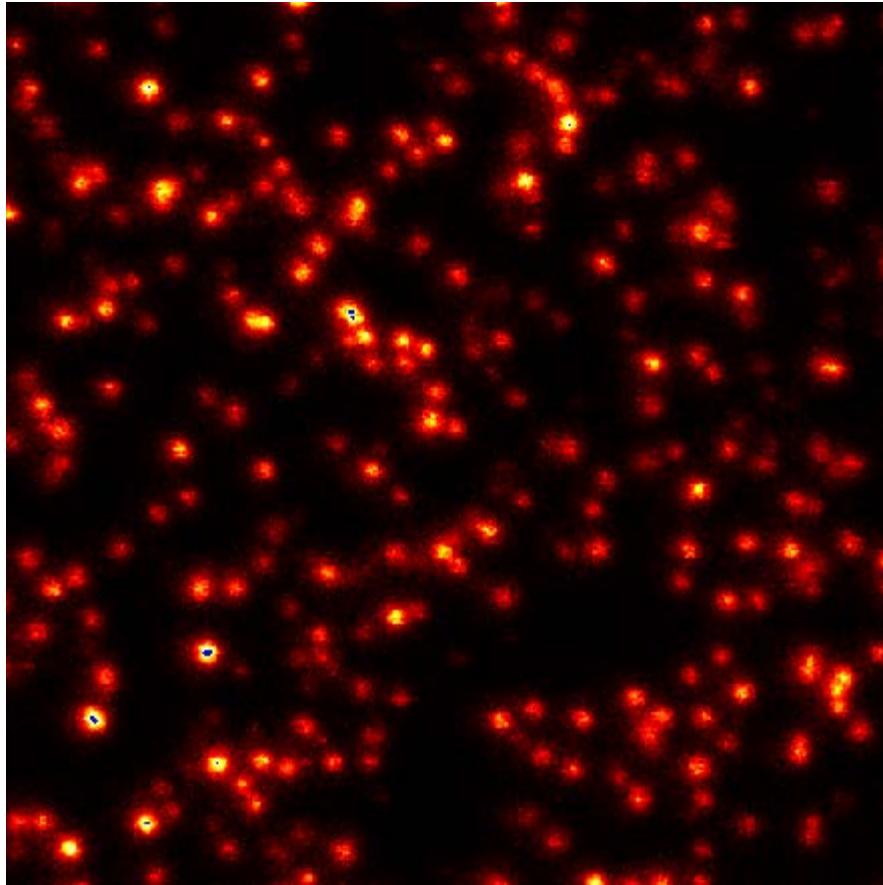


20nm Crimson beads
633nm exc, 90ps, 30kW/cm²
785nm STED 200ps, 76MHz

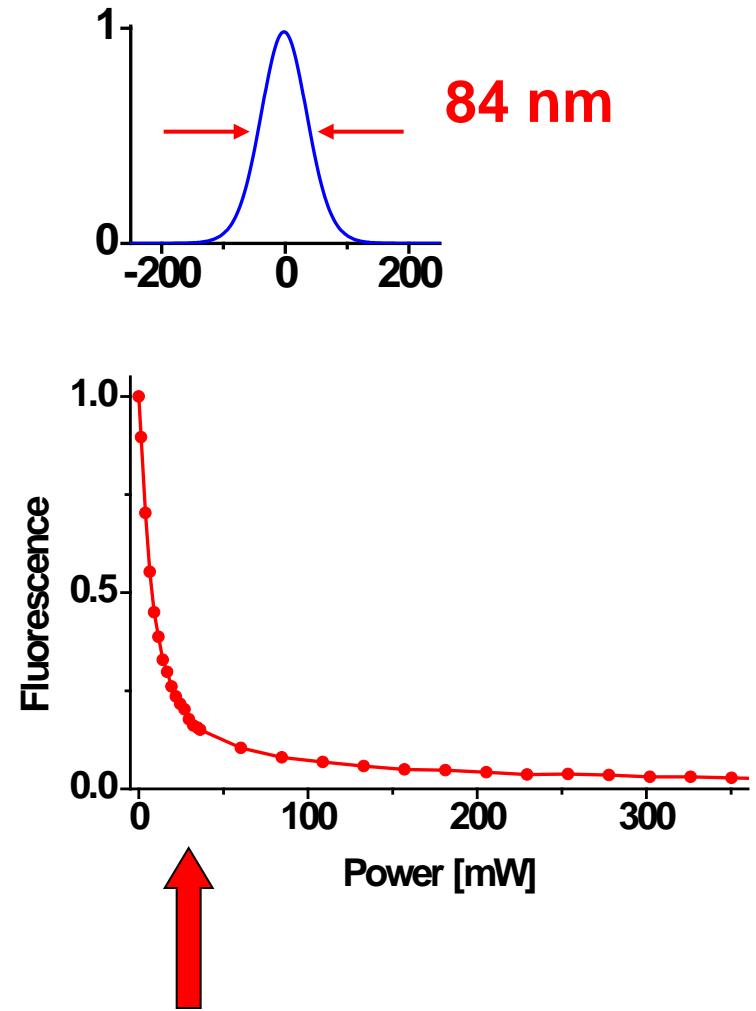


STED-Microscopy

Sub-Diffraction Imaging

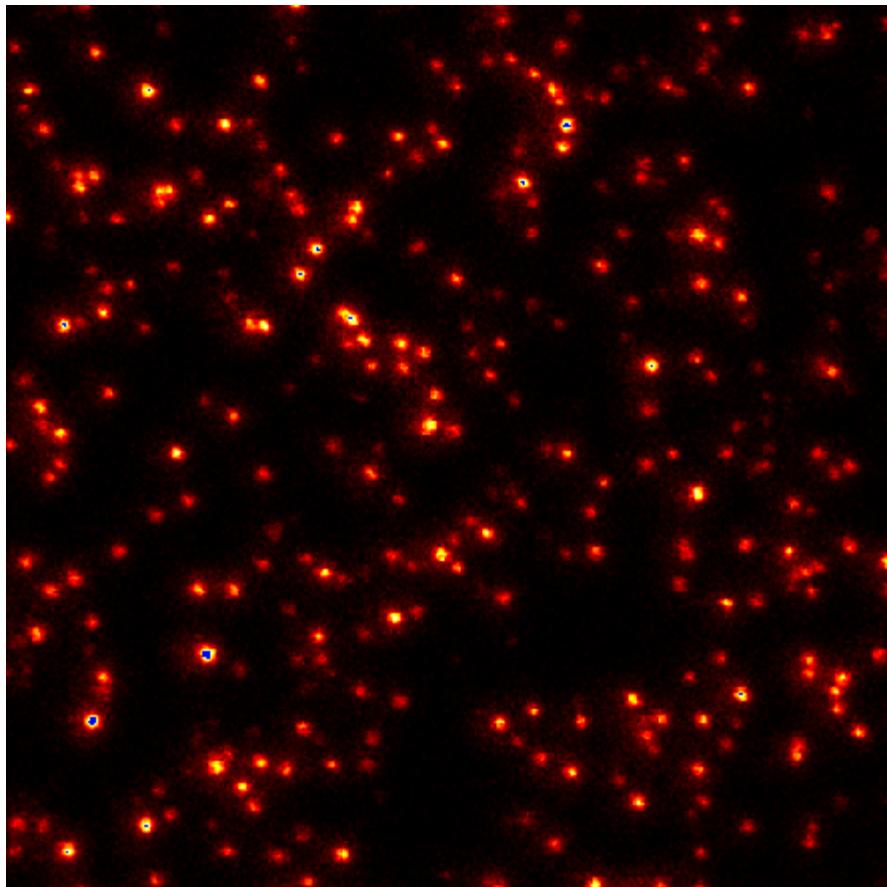


20nm Crimson beads
633nm exc, 90ps, 30kW/cm²
785nm STED 200ps, 76MHz

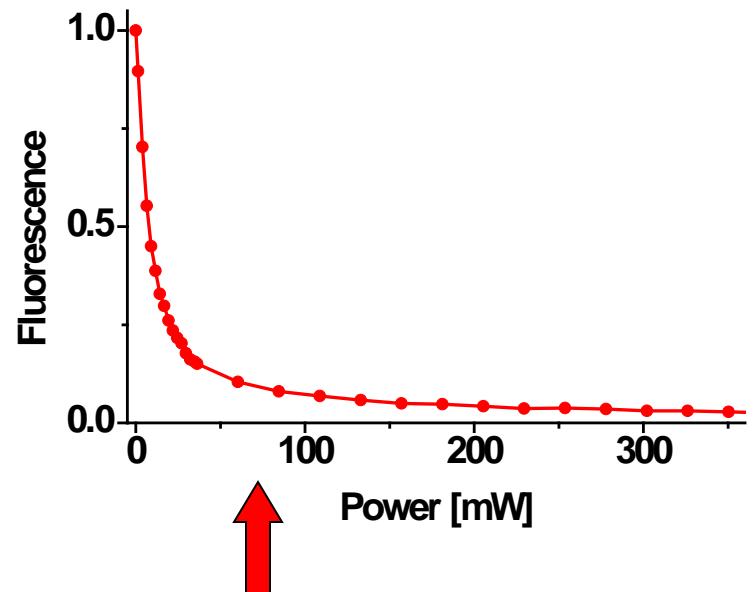
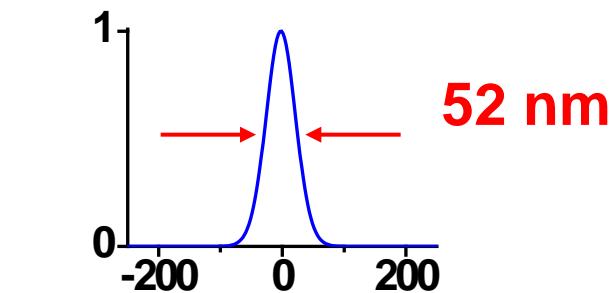


STED-Microscopy

Sub-Diffraction Imaging

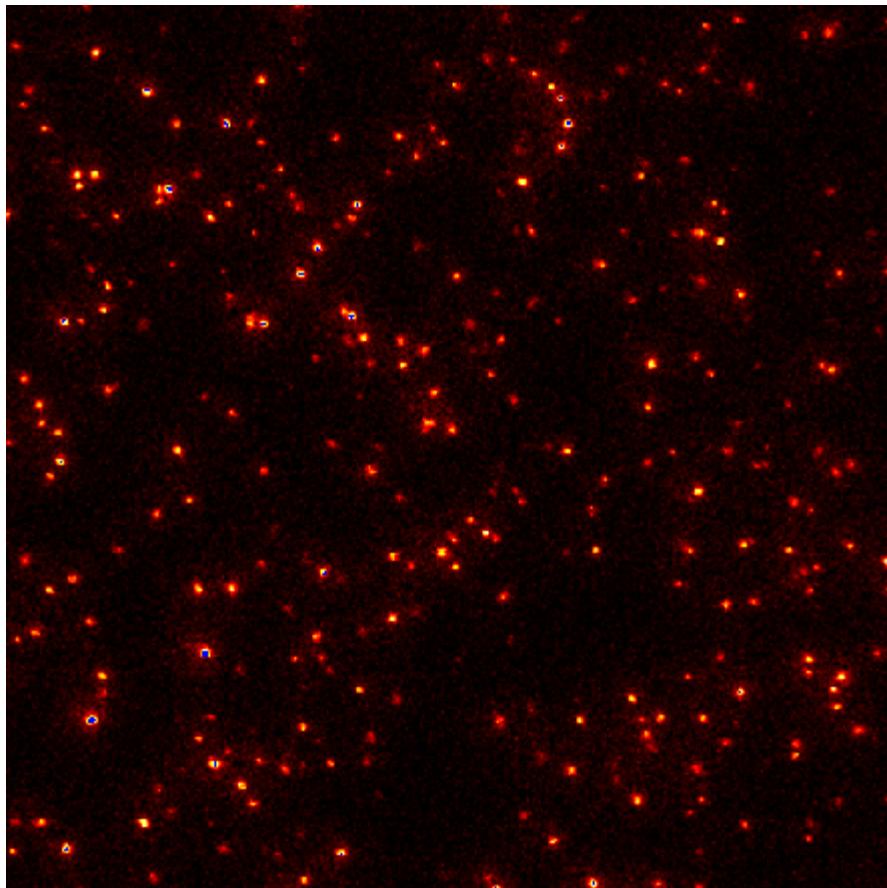


20nm Crimson beads
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785nm STED 200ps, 76MHz

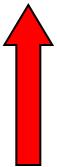
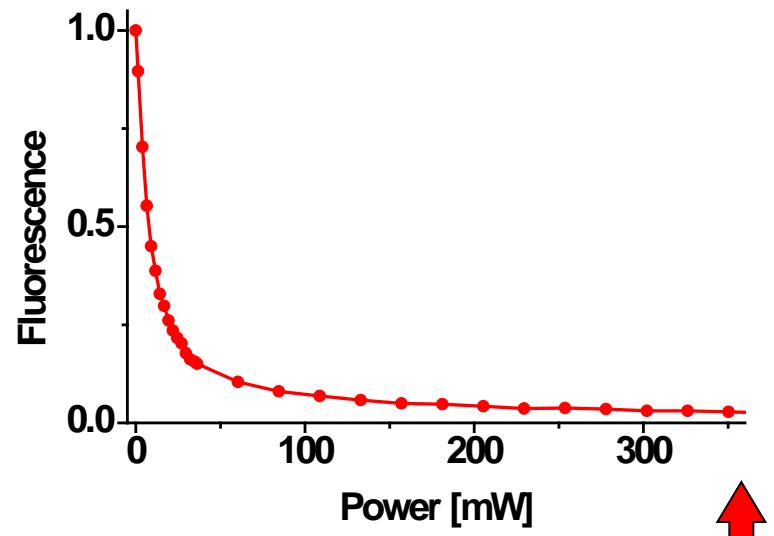
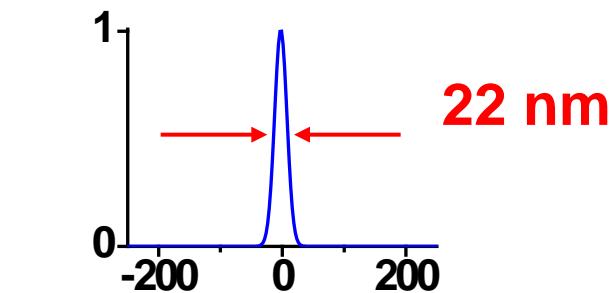


STED-Microscopy

Sub-Diffraction Imaging

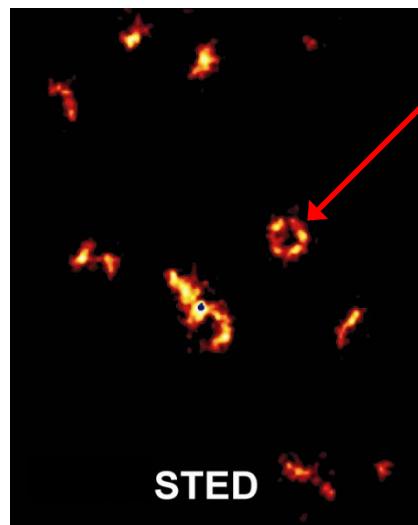
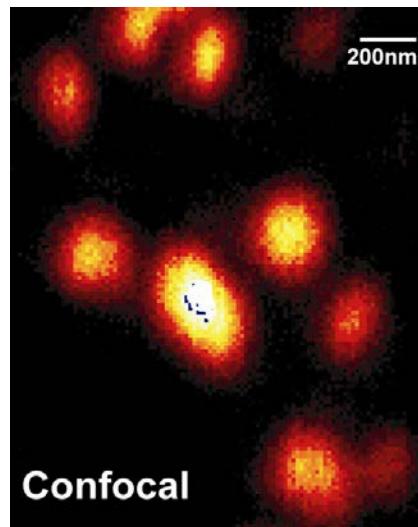


20nm Crimson beads
633nm exc, 90ps, 30kW/cm²
785nm STED 200ps, 76MHz



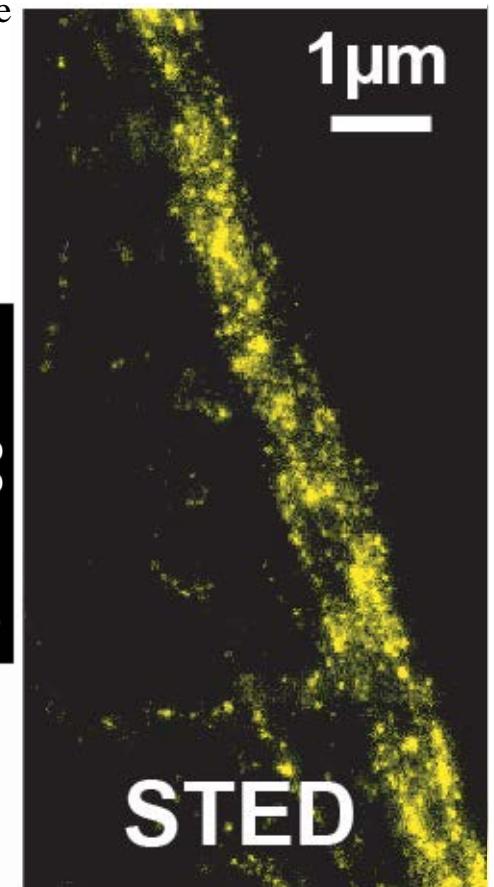
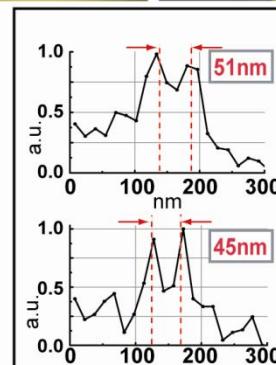
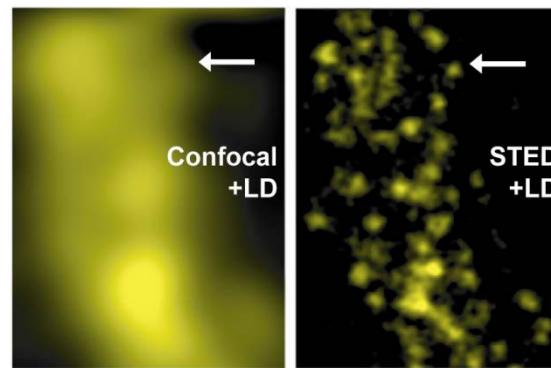
STED Microscopy

Cellular Imaging



synaptic proteins on endosomes
of PC 12 cells
(neuroendocrine activity;
generate synaptic vesicles)
Atto532-synaptophysin
LD

protein-heavy subunit of neurofilaments
in the human neuroblastoma cell line
SH-SY5Y (retinoic acid–BDNF-
differentiated);
establishes cross-links to organize
and stabilize neurofilaments in axons



STED Microscopy

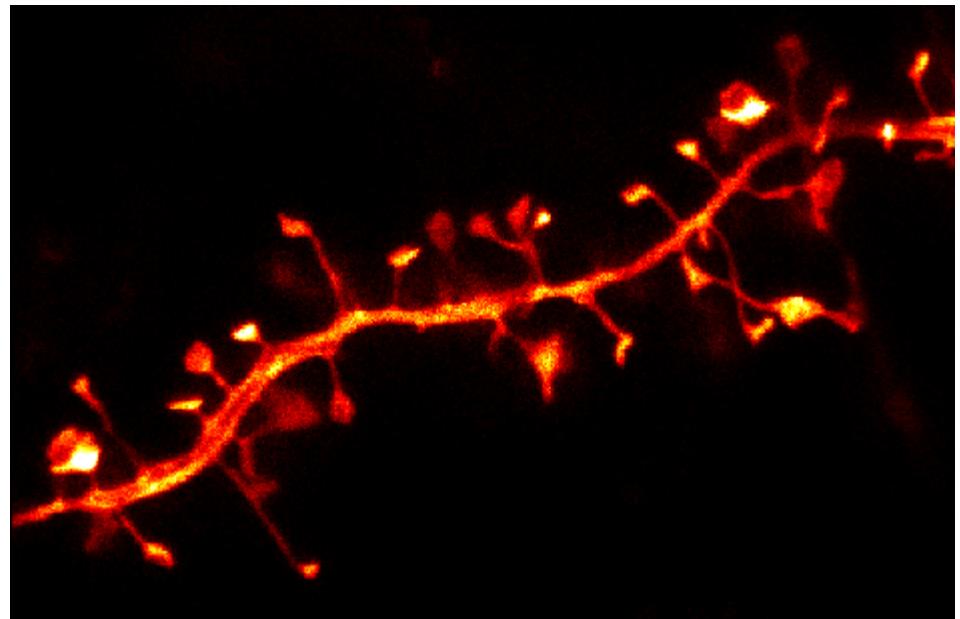
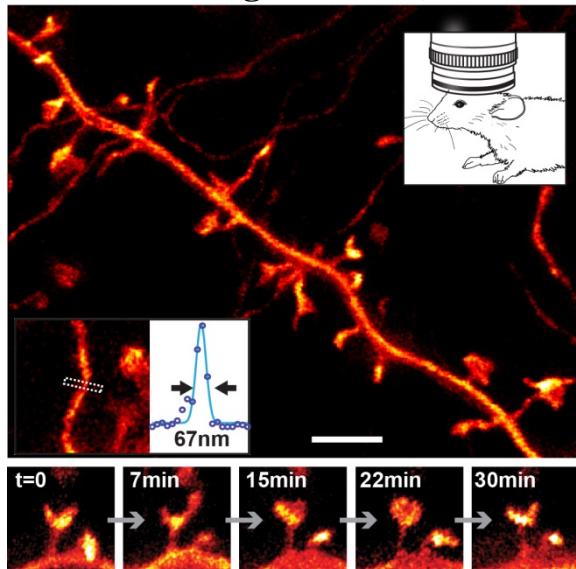
Inside Living Cells - Dynamics

YFP-transgenic mouse
Hippocampal slice
CA1 neuron
(PNAS Nägerl et al 2008)
(BiophysJ 2011)

Live Mouse

YFP

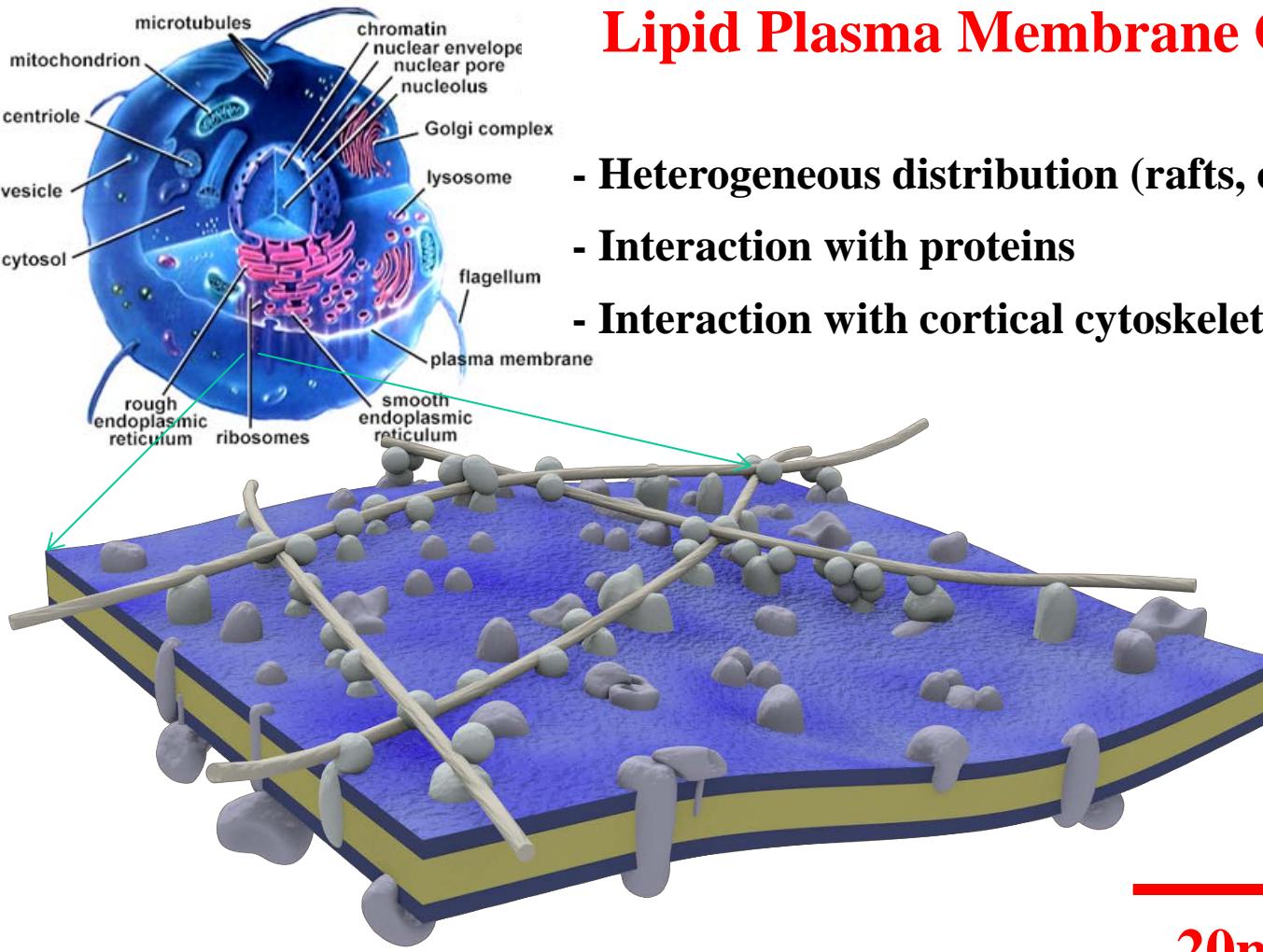
(Science Berning et al 2012)



Live-Cell (inside)
Multi-Color (more complex)
Two-Photon excitation
3D possible
Conventional dyes, GFP, ...

Lipid Plasma Membrane Organization

Nanoscale



Lipid Plasma Membrane Organization:

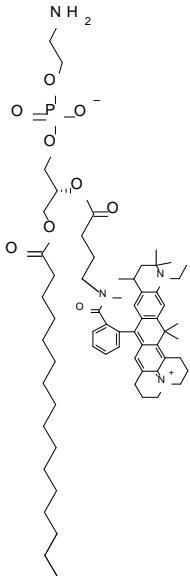
- Heterogeneous distribution (rafts, curvature, ...)
- Interaction with proteins
- Interaction with cortical cytoskeleton

—
20nm
Small spatial scales!!!!

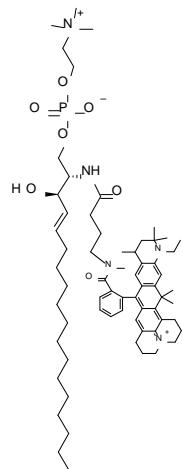
Lipid Plasma Membrane Organization

Fluorescence Recordings: Lipids

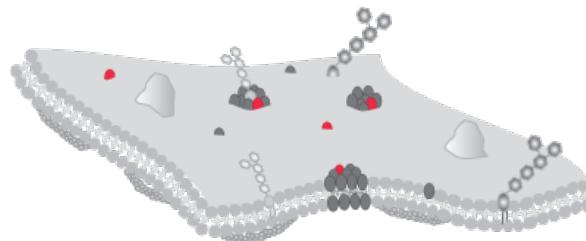
Phosphoglycerolipid:
Atto647N-phosphoethanolamine (PE)



Sphingolipid:
Atto647N-sphingomyelin (SM)



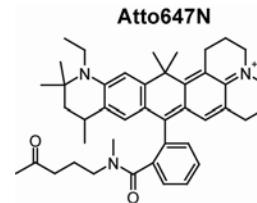
Live PtK2 cells:
physiological conditions
incorporation in plasma membrane



BSA
complex

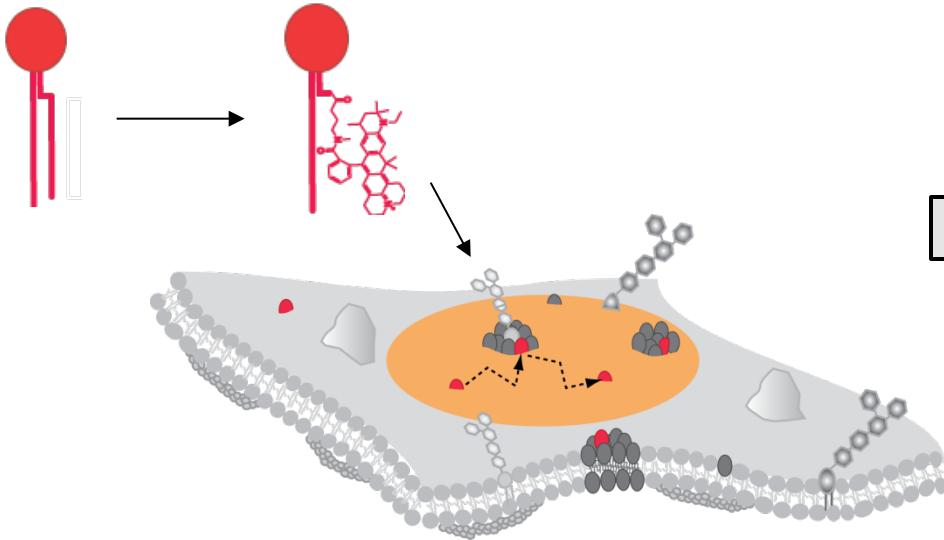


Labeling

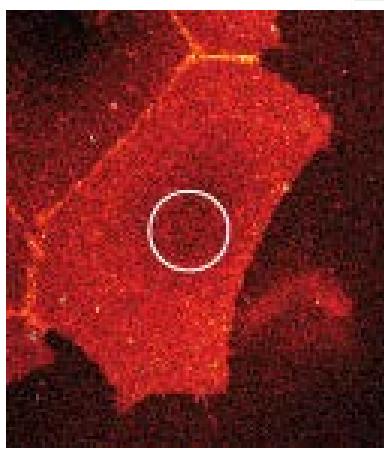


Lipid Plasma Membrane Organization

Confocal Recordings

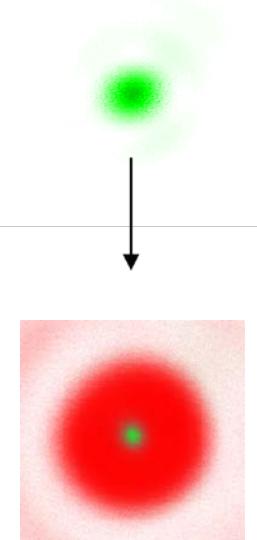
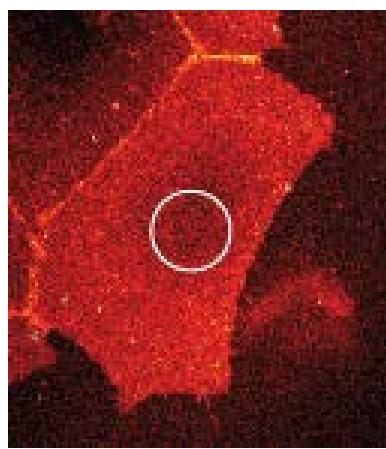
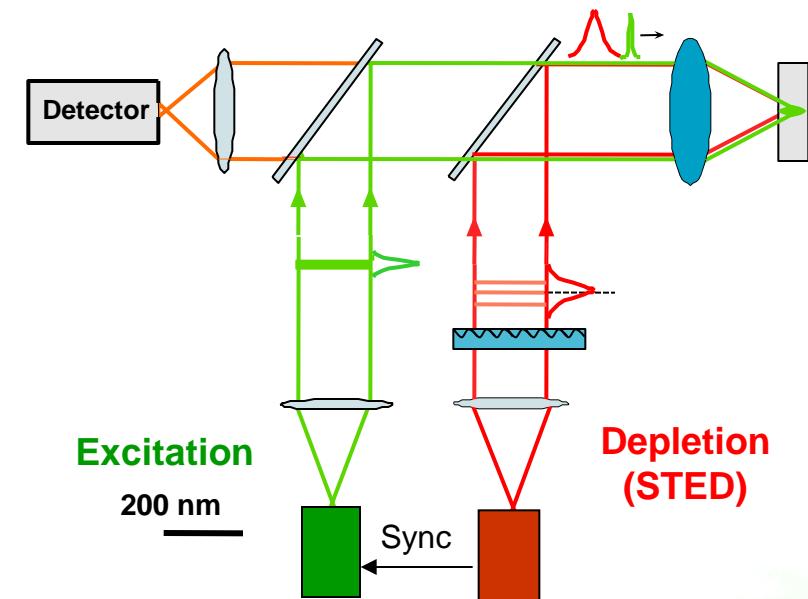
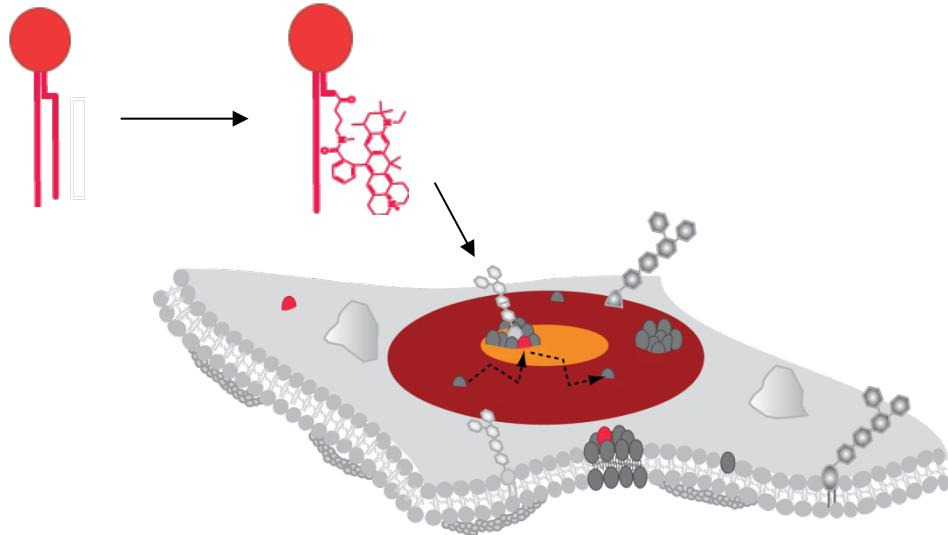


Homogeneous distribution



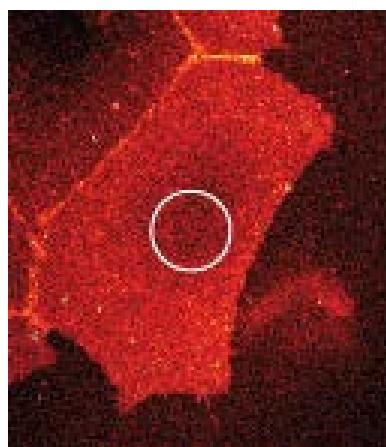
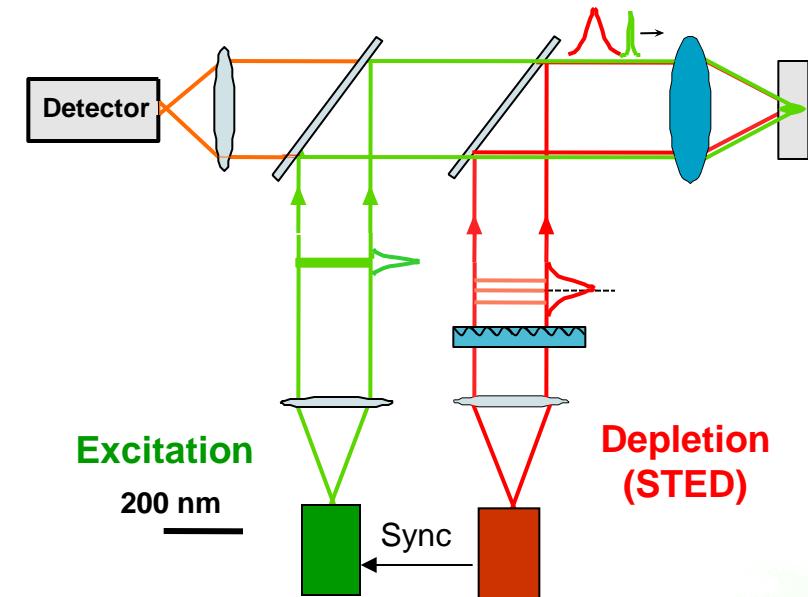
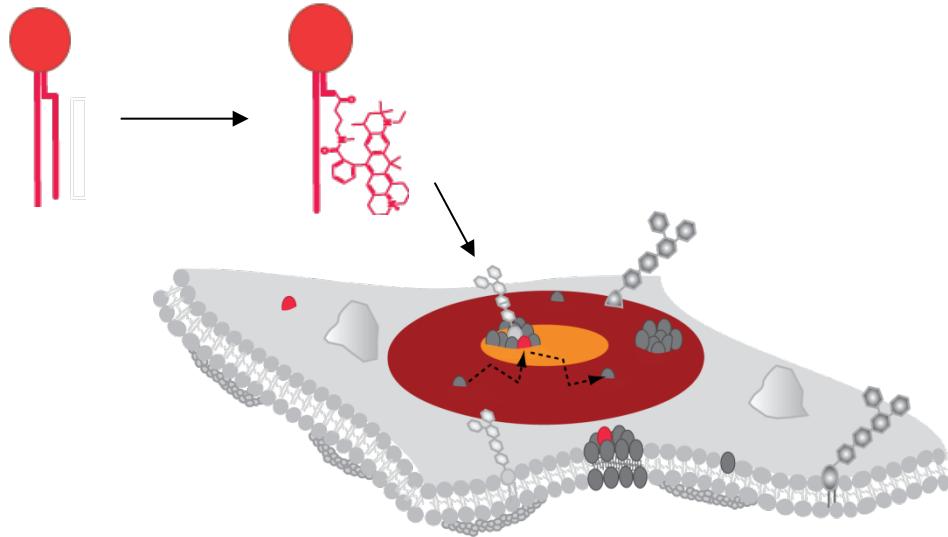
Lipid Plasma Membrane Organization

STED Nanoscopy Measurement



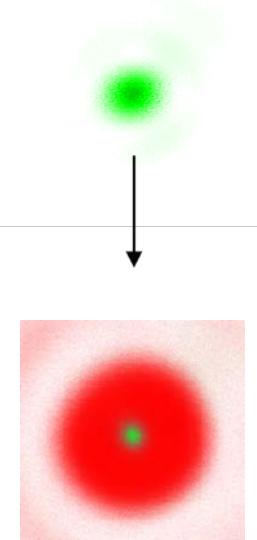
Lipid Plasma Membrane Organization

STED Nanoscopy Measurement



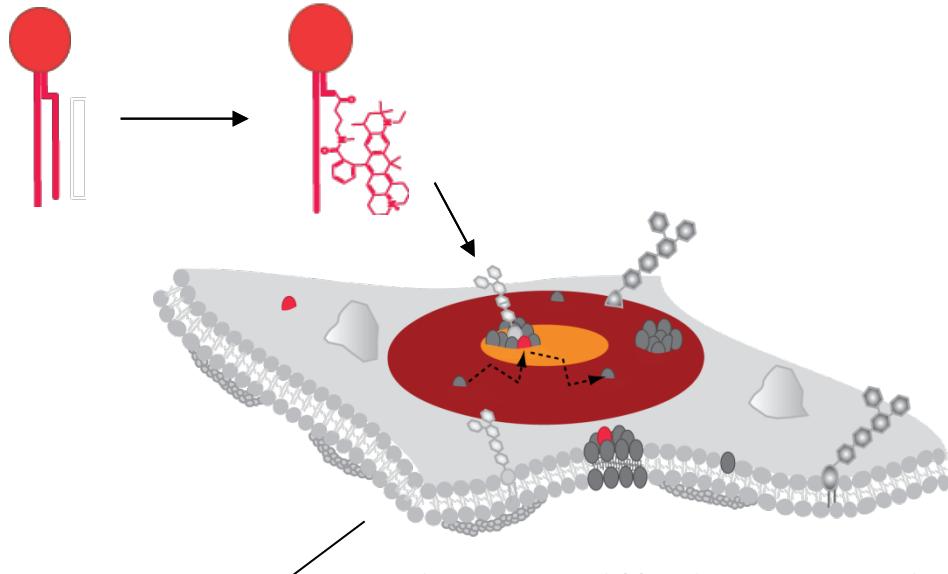
Homogeneous distribution

Fast diffusion → Limited temporal resolution!

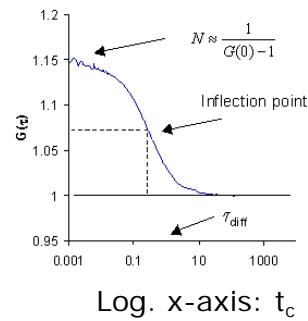
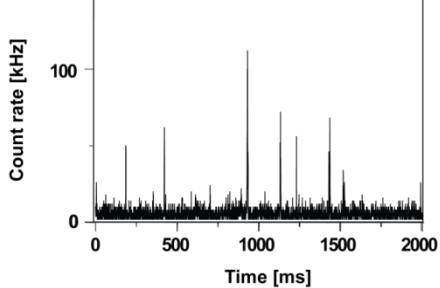


Lipid Plasma Membrane Dynamics

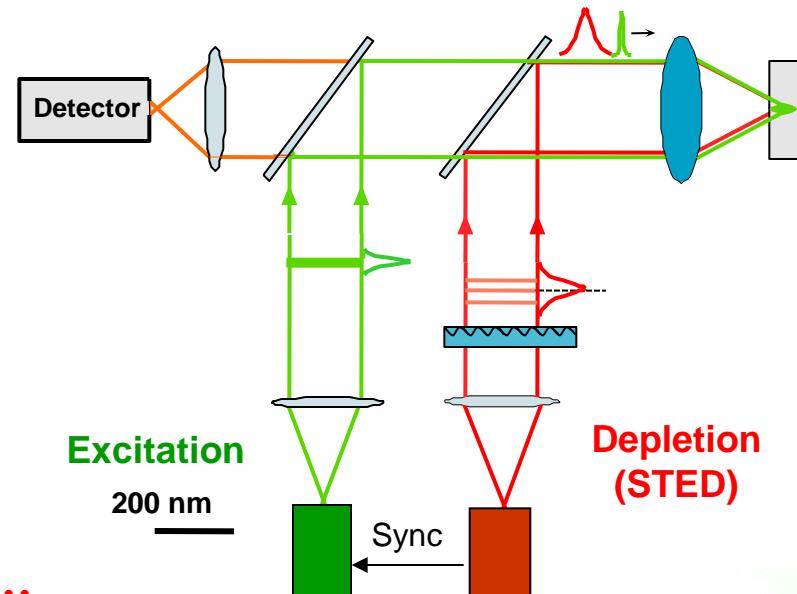
STED Nanoscopy Measurement



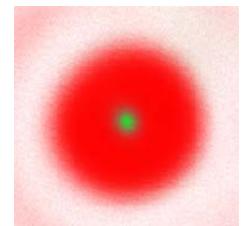
Discover diffusion dynamics!!!
Fluorescence Correlation Spectroscopy (FCS)



Eggeling et al Nature 2009



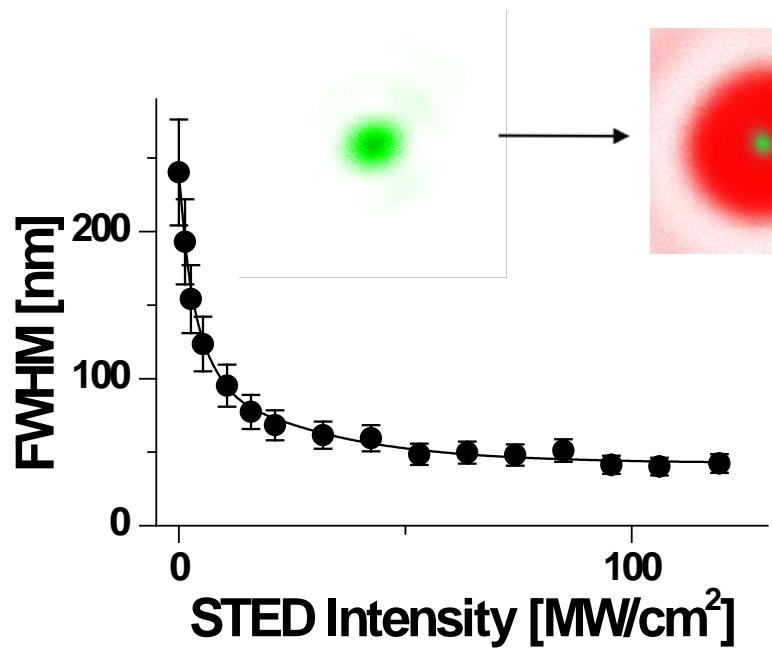
molecular diffusion coefficient
=
molecular mobility



Live Cell Nanoscopy

STED-FCS

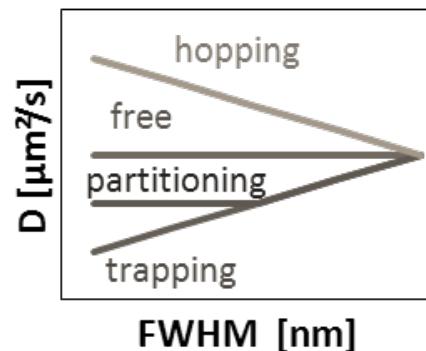
STED-Microscopy: Tuning of observation area



STED-FCS
Determine transit time
for different sizes of observation areas
(different STED intensities)

Calculate
apparent diffusion coefficient:
 $D \sim \text{area} / \text{transit time}$

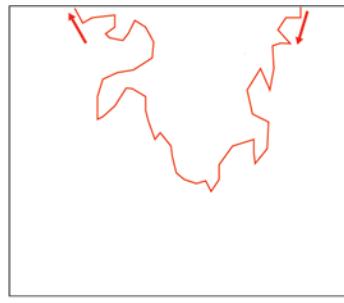
Dependencies: $D(\text{diameter})$
 $240\text{nm} \rightarrow 30/40\text{nm}$
Varies for different diffusion modes



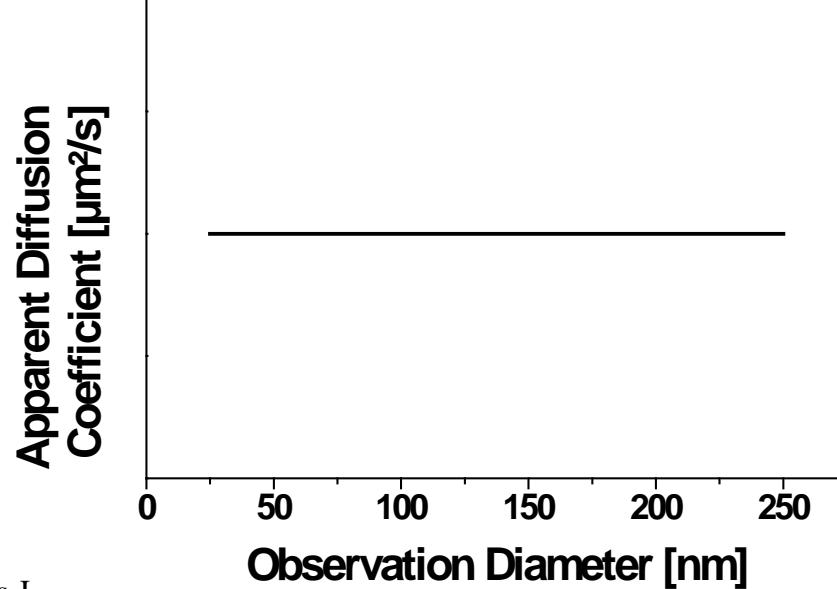
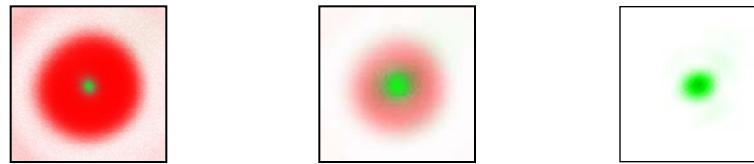
Live Cell Nanoscopy

STED-FCS - Diffusion Models

Free diffusion



← STED Intensity



Wawrzynieck et al. Biophys J.

2005 December; 89(6)

Eggeling et al. Nature 457,
1159-1162 ,2009

Mueller et al. Biophys J 2011

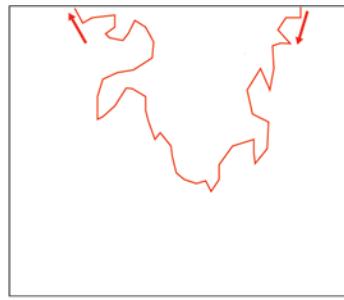
Apparent diffusion coefficient:

$D \sim \text{area} / \text{transit time}$

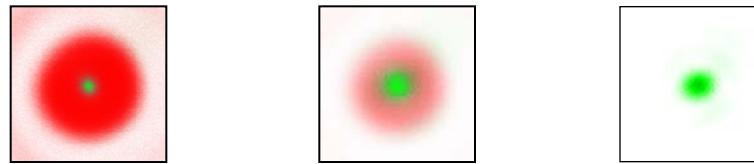
Live Cell Nanoscopy

STED-FCS - Diffusion Models

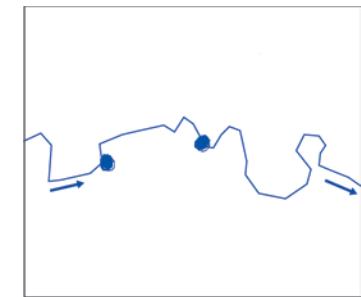
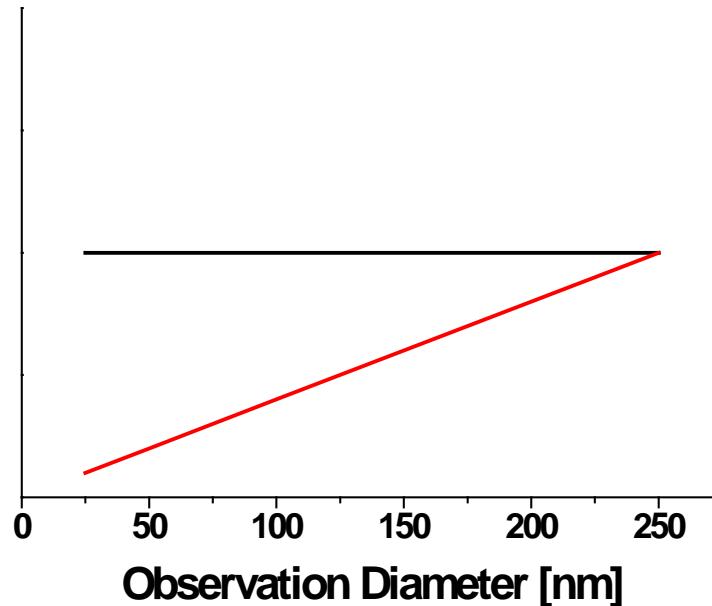
Free diffusion



← STED Intensity



Apparent Diffusion
Coefficient [$\mu\text{m}^2/\text{s}$]



Trapping

Wawrzinek et al. Biophys J.

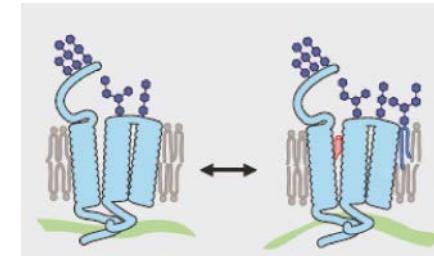
2005 December; 89(6)

Eggeling et al. Nature 457,
1159-1162 ,2009

Mueller et al. Biophys J 2011

Apparent diffusion coefficient:

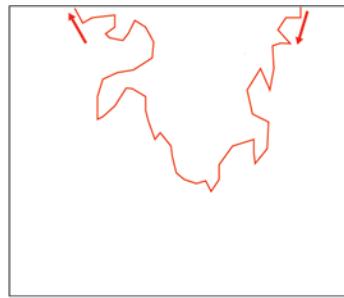
$D \sim \text{area} / \text{transit time}$



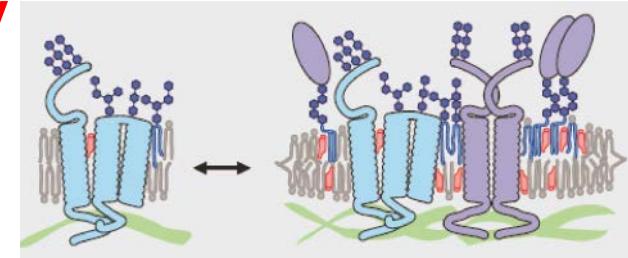
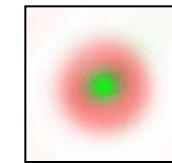
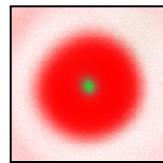
Live Cell Nanoscopy

STED-FCS - Diffusion Models

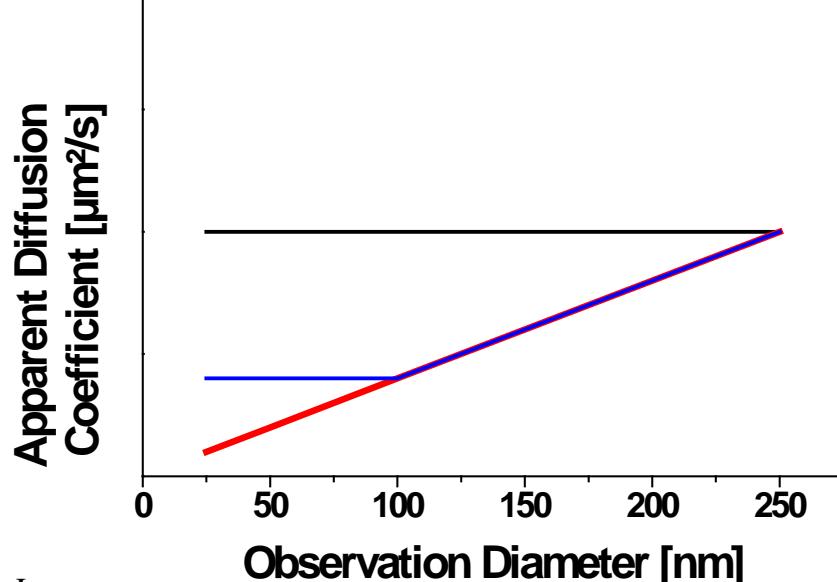
Free diffusion



← STED Intensity



Domain incorporation



Wawrzynieck et al. Biophys J.

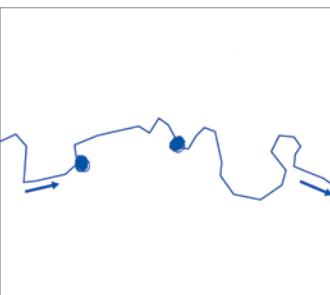
2005 December; 89(6)

Eggeling et al. Nature 457,
1159-1162 ,2009

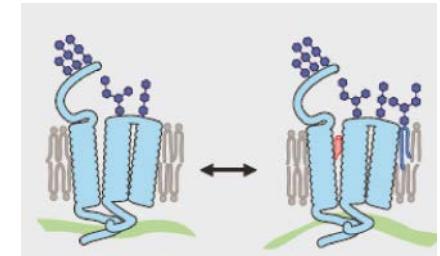
Mueller et al. Biophys J 2011

Apparent diffusion coefficient:

$D \sim \text{area} / \text{transit time}$

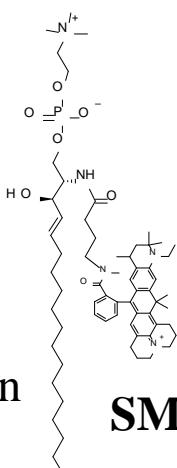
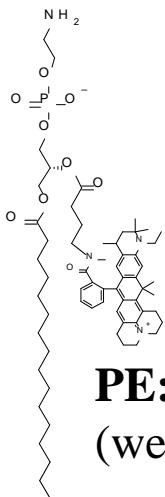
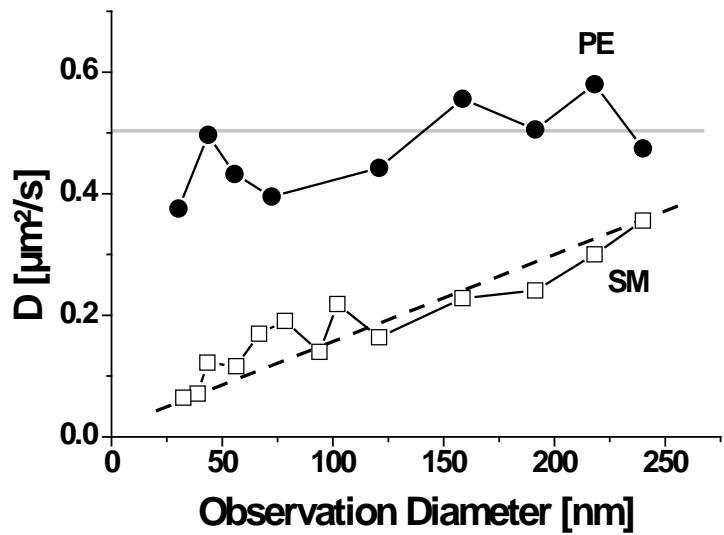


Trapping

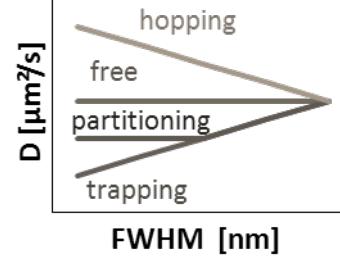
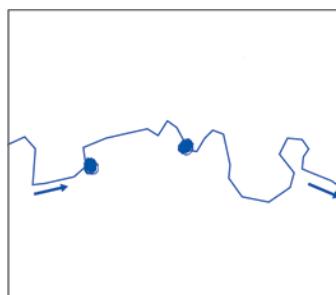


STED-FCS

Lipid Membrane Diffusion + Interactions: PE + SM

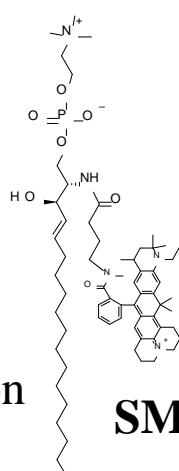
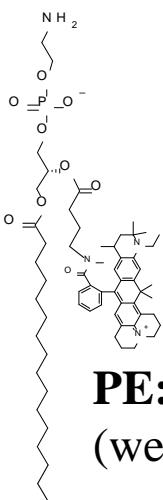
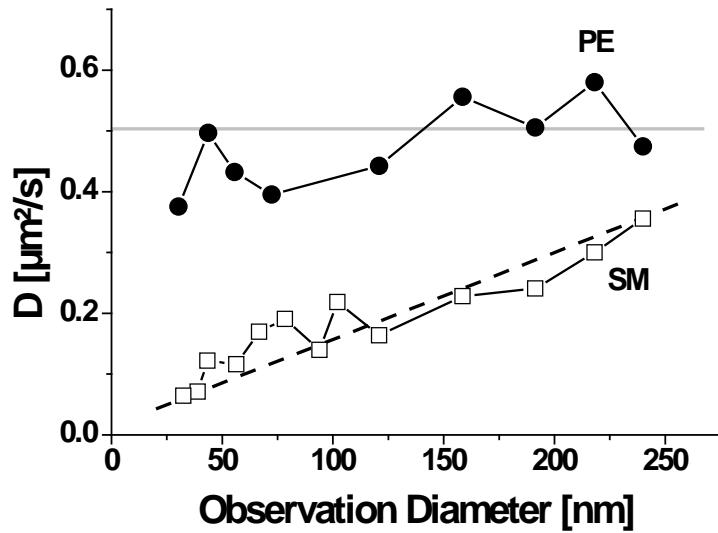


Eggeling et al. *Nature* 2009
Mueller et al. *Biophys J* 2011



STED-FCS

Lipid Membrane Diffusion + Interactions: PE + SM



→ Complex on molecular scale
(proteins, lipid-shells, ...)

~10 ms, no movement during trapping

Cholesterol-assisted
(COase/β-Cyclo-Dextrin/Zaragozic acid...)

Binding partner bound to cytoskeleton
(Latrunculin/Jasplakinolide/Nocodazole...)

Slight dependence on endogenous SM level
(Myriocin)

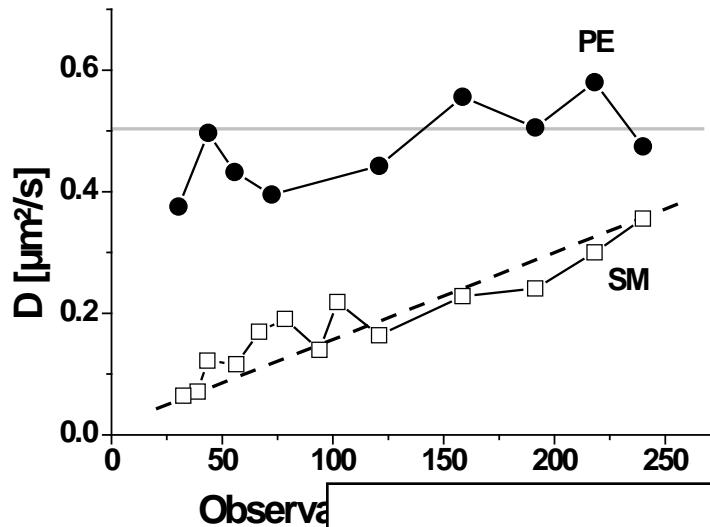
Dependence on lipid structure
(but not dye)

Eggeling et al. *Nature* 2009
Mueller et al. *Biophys J* 2011



STED-FCS

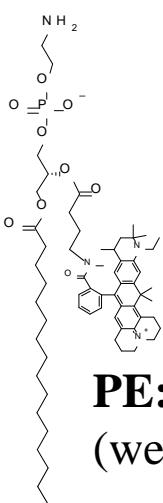
Lipid Membrane Diffusion + Interactions: PE + SM



→ Complex on molecular scale
(proteins, lipid-shells, ...)

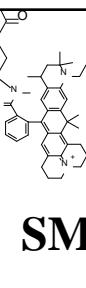
~10 ms, no movement during trapping

Cholesterol-assisted
(COase/β-Cyclo-Dextrin/Zaragozic acid...)



PE: free diffusion
(weak trapping)

STED-FCS:
New approach to study molecular interactions!



SM: trapping

keleton
(Nocodazole...)

SM level

Dependence on lipid structure
(but not dye)

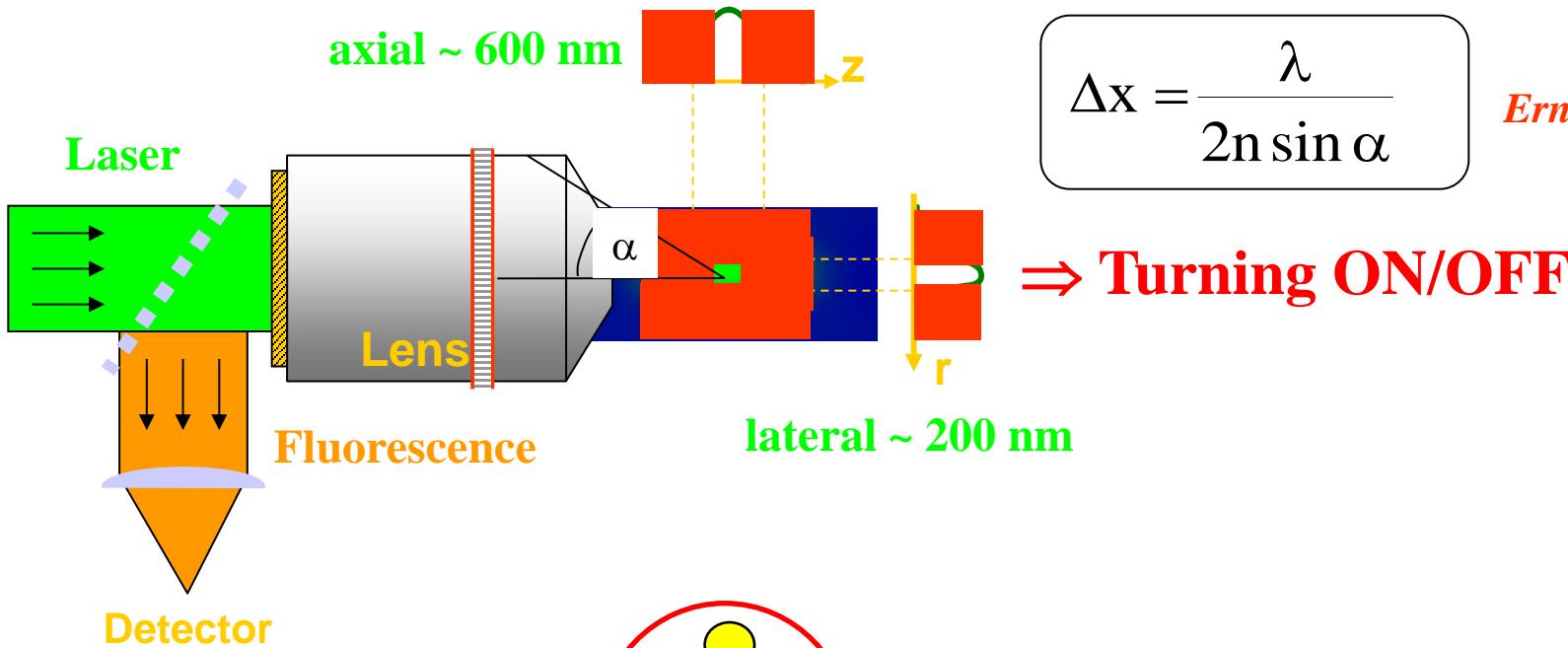
Eggeling et al. *Nature* 2009
Mueller et al. *Biophys J* 2011



FWHM [nm]

Far-Field Microscopy

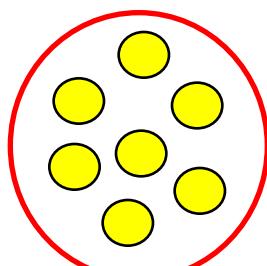
Surpassing the Resolution Limit: Turning ON/OFF



$$\Delta x = \frac{\lambda}{2n \sin \alpha}$$

Ernst Abbe 1873

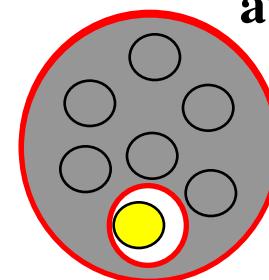
⇒ Turning ON/OFF



⇒ Observation area / Resolution

lateral << 200 nm
axial << 600 nm

Turn OFF all objects but
at a single point



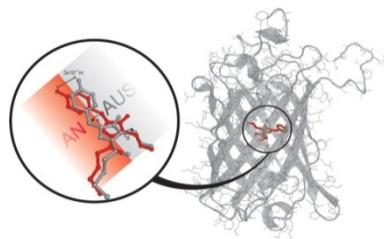
STED/
RESOLFT

Far-Field RESOLFT Nanoscopy

Reversibly Photoswitchable Fluorescent Proteins

Photoswitchable Proteins

ON \longleftrightarrow OFF

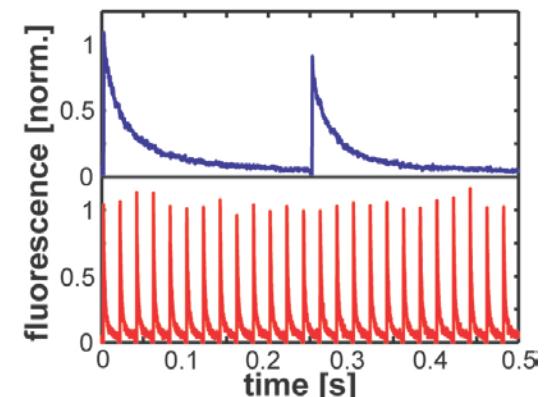


Photoisomerisation
cis-trans conformational states
dark (trans)- bright (cis)
Andresen et al. (2005) PNAS

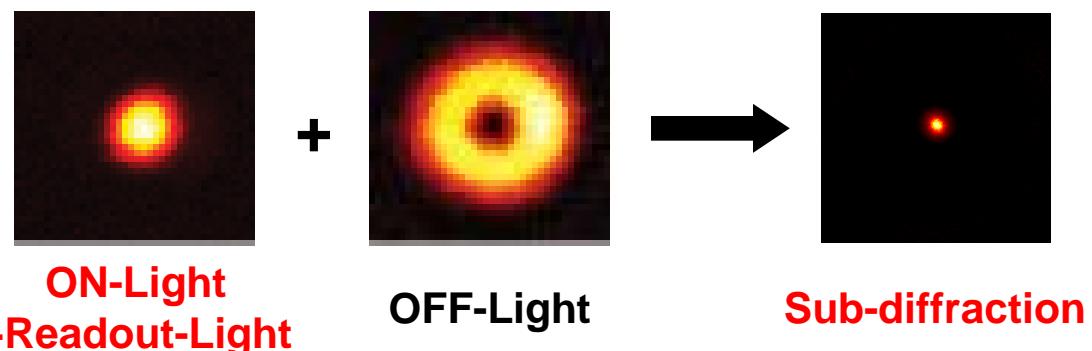
Dronpa \rightarrow rsFastLime
Stiel et al, Biochem J 2007

GFP \rightarrow photoswitching (rsEGFP)
Grotjohann et al, Nature 2011

Switch-off + Readout: 488nm
Switch-on: 405nm



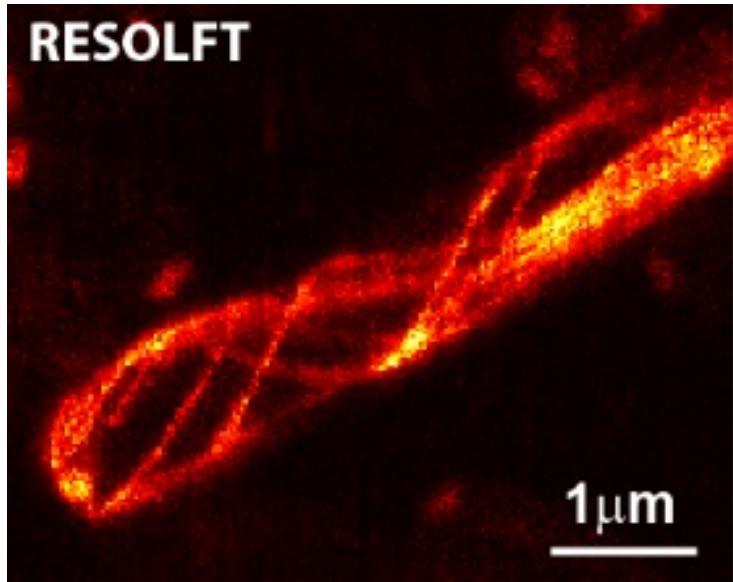
ON/OFF at low CW powers
 $nW - \mu W (\sim kW/cm^2)$



RESOLFT = Reversible Saturable Optical Fluorescence Transition

Far-Field RESOLFT Nanoscopy

Reversibly Photoswitchable Fluorescent Proteins



Excellent for Live-Cell (low light levels)

Multi-Color (new fluorescent proteins)
3D possible
Photoswitchable proteins / dyes

Intensity $\approx 1 \text{ kW/cm}^2$

Citrine \rightarrow Dreiklang

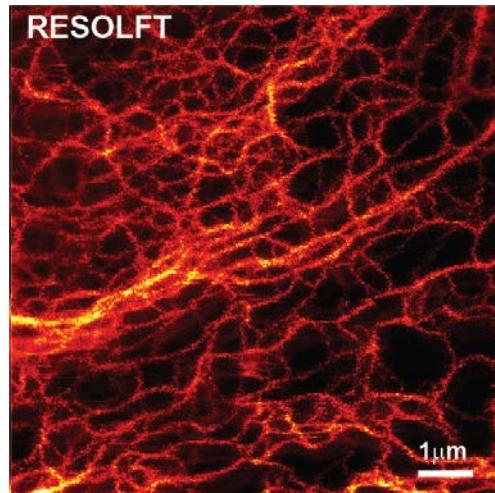
Brakemann et al, Nature Biotechnol. 2011

Switch-on: 405 nm

Switch-off: 355 nm

Read-out: 488 nm

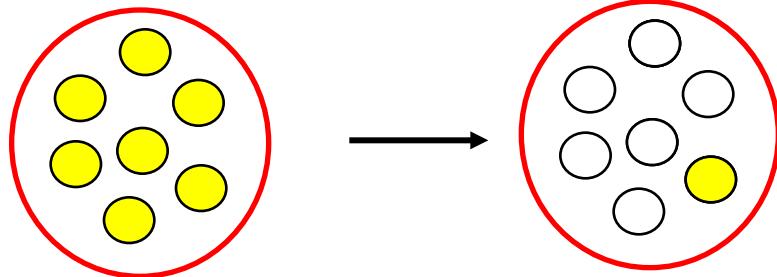
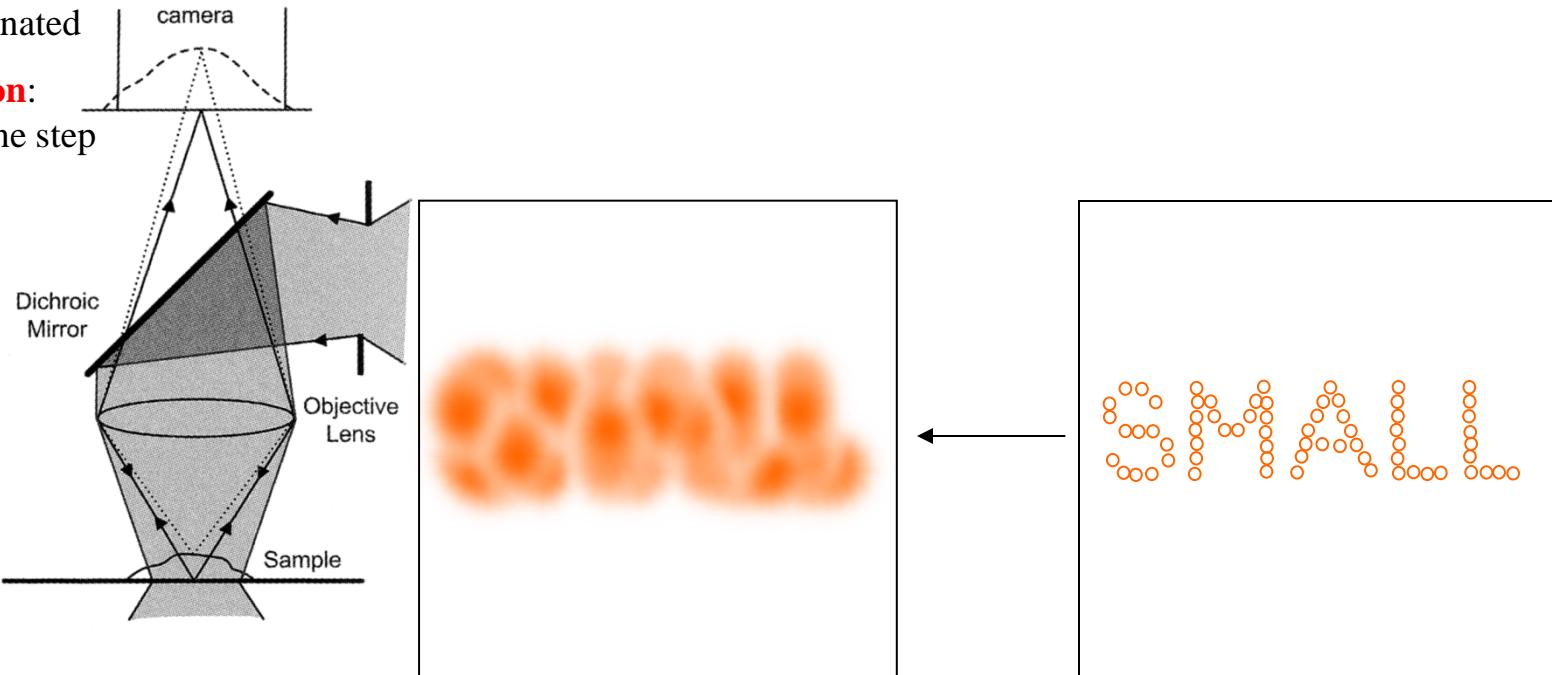
Keratin19-Dreiklang
expressed
in living PtK2 cells



(d)STORM/(f)PALM Far-Field Nanoscopy

Resolution Limit – Camera Detection

- Large area illuminated
- **Camera detection:**
image taken in one step

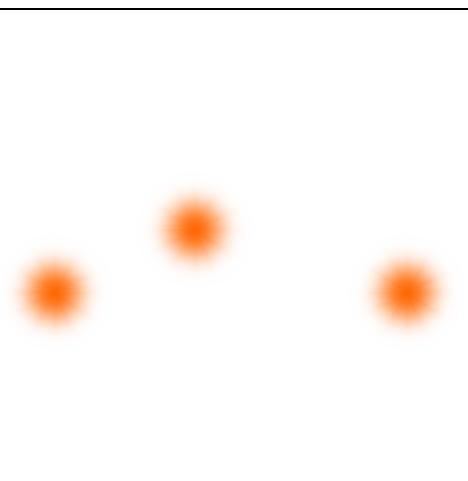


**Turn OFF all but
a single object**

**(d)STORM/
(f)PALM**

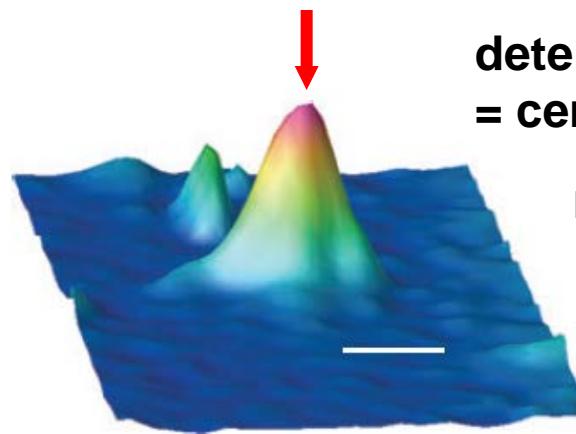
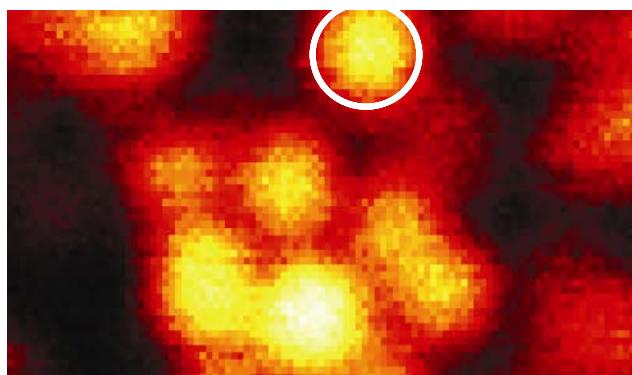
Far-Field Nanoscopy

ON/OFF: Single-Molecule



Localization of single molecules
- accuracy down to the molecular scale

Diffraction-limited



determination of exact position
= center of diffr.-lim. peak

N_{photons} = number of photons detected
per single molecule

$$\Delta x = \Delta x_{\text{diffr.}} N_{\text{photons}}^{0.5}$$

1000 photons: ~ 10nm

Moerner, Nat. Meth. 2006

Far-Field Nanoscopy

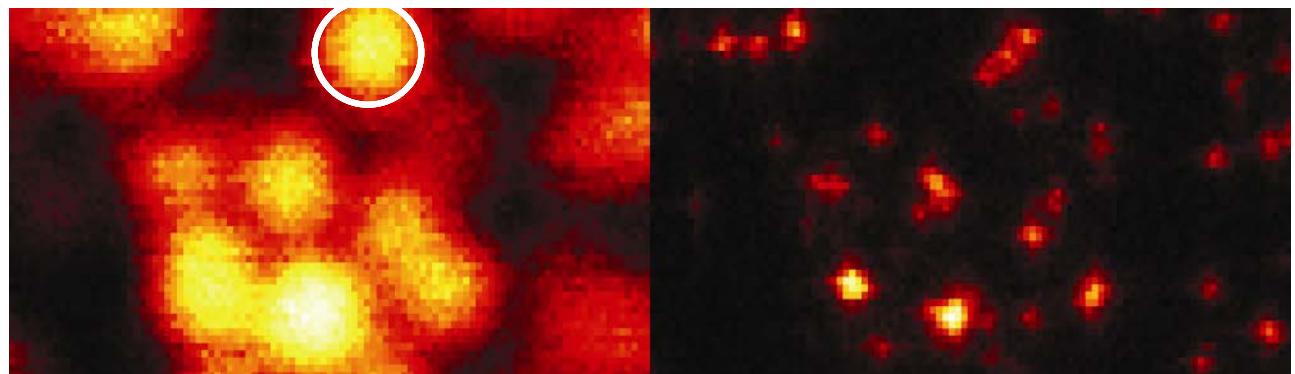
ON/OFF: Single-Molecule

SMALL

Localization of single molecules

- accuracy down to the molecular scale

Diffraction-limited



⇒ really single molecule?

(localization not resolution)

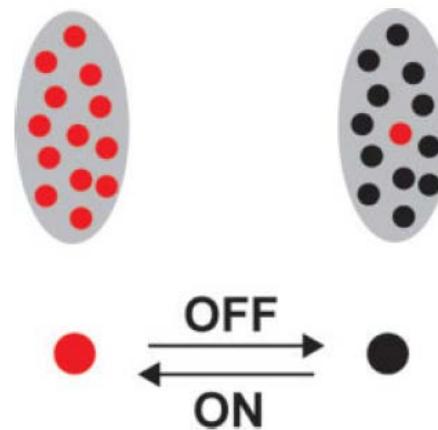
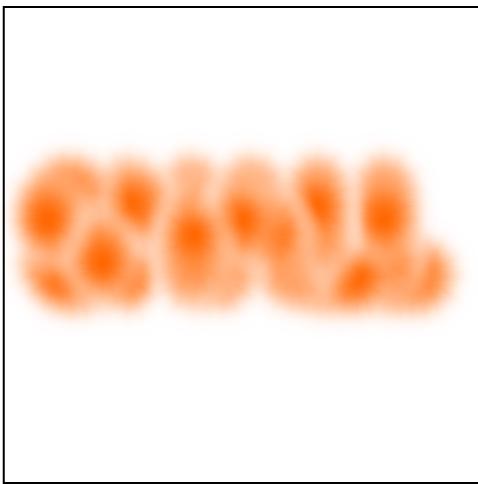


Far-Field Nanoscopy

Single-Molecule Switching



Downscale ensemble to
single isolated molecules



⇒ Single molecules separated by more
than the diffraction limit

⇒ Localization of real single molecule

Far-Field Nanoscopy

Single-Molecule Switching + Localization



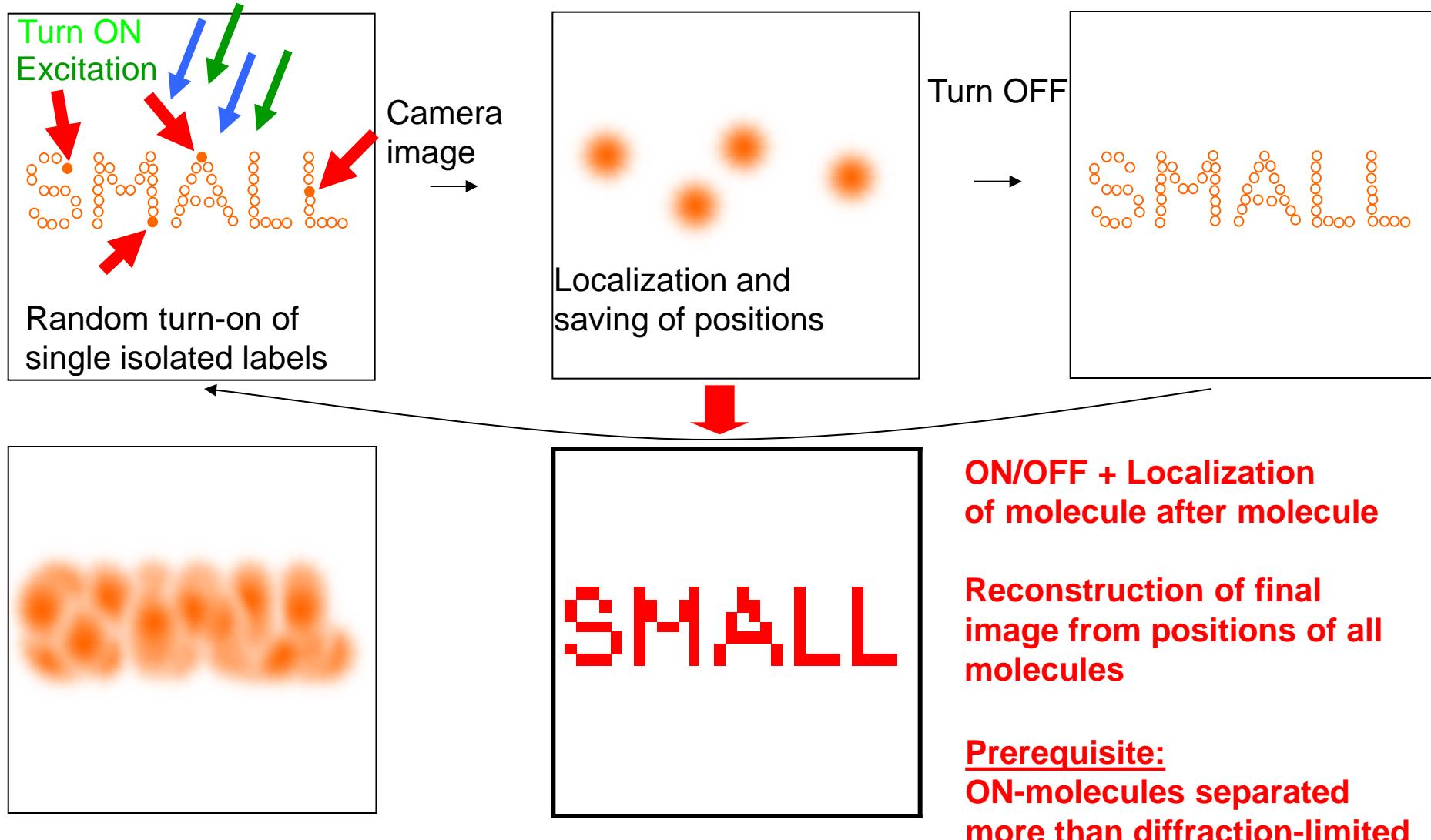
Far-Field Nanoscopy

Single-Molecule Switching + Localization



Far-Field Nanoscopy

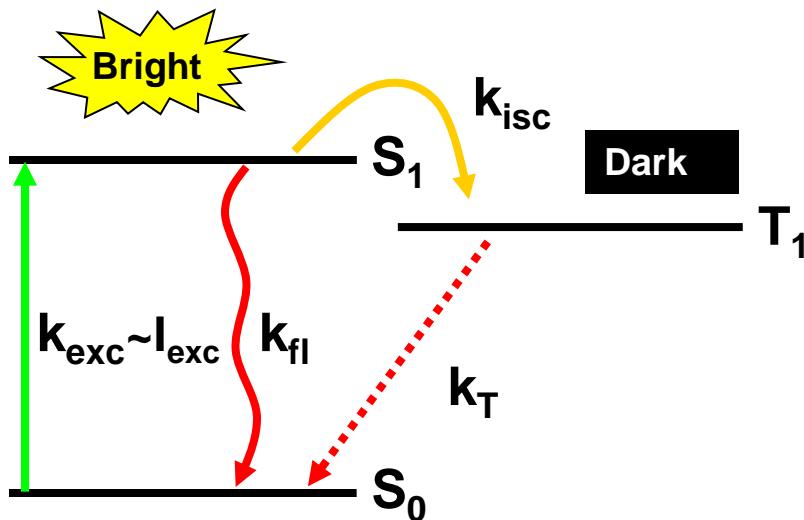
Single-Molecule Switching + Localization



Far-Field Nanoscopy

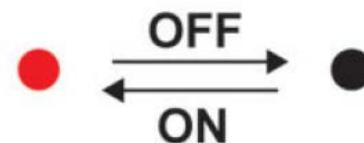
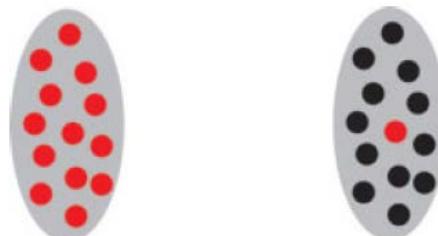
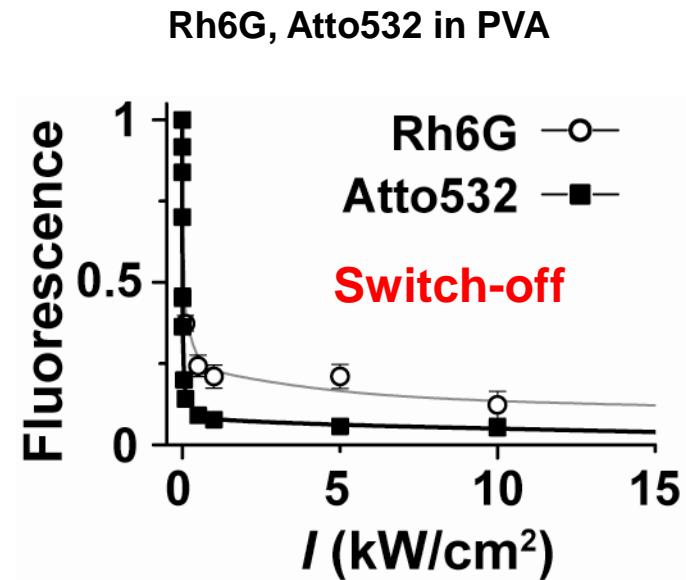
*GSDIM – Ground State Depletion microscopy
followed by Individual Molecule return*

Turn OFF:
Shelve into long-lived dark triplet state



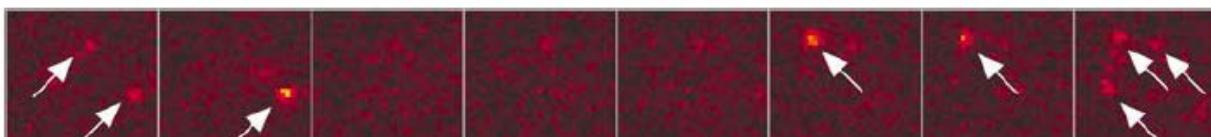
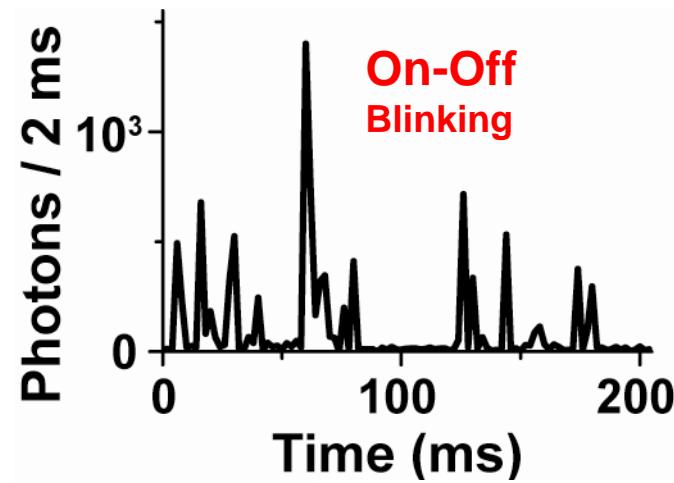
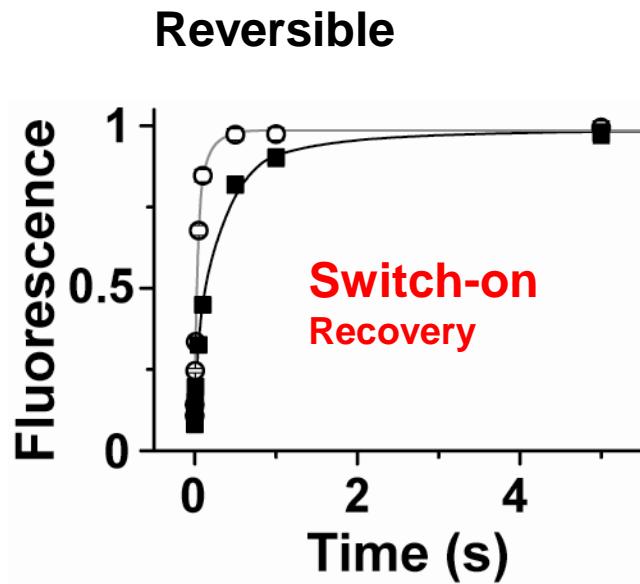
Conventional fluorophores have dark state!!

Downscale ensemble
to isolated
single- molecule

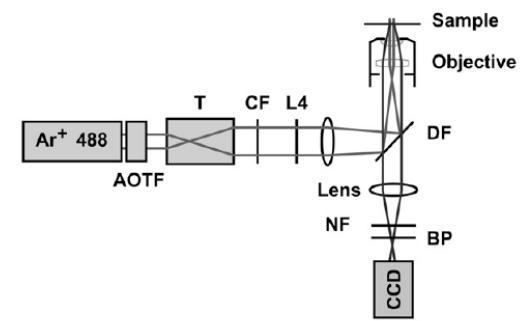


Far-Field Nanoscopy

GSDIM

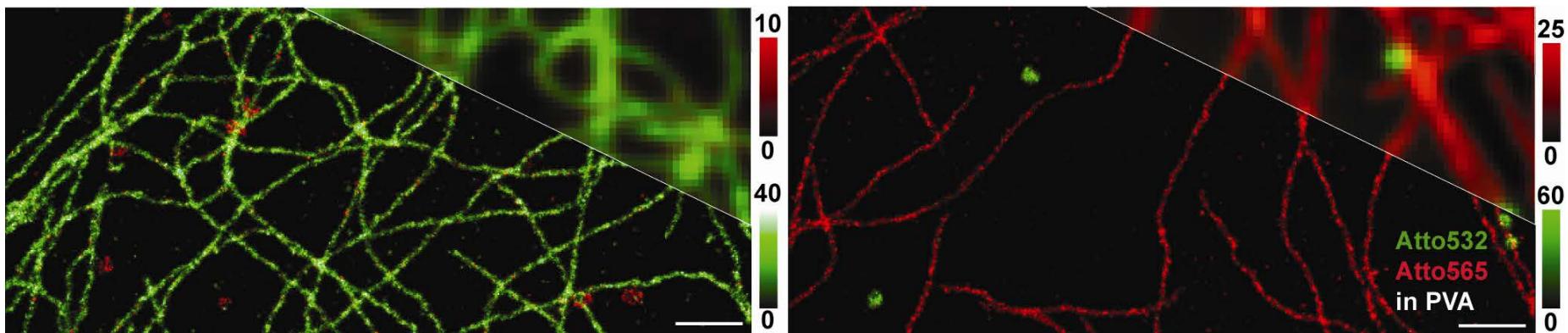


$\Delta x < 20\text{nm}$ ($N \sim 1000$)



Far-Field Nanoscopy

GSDIM

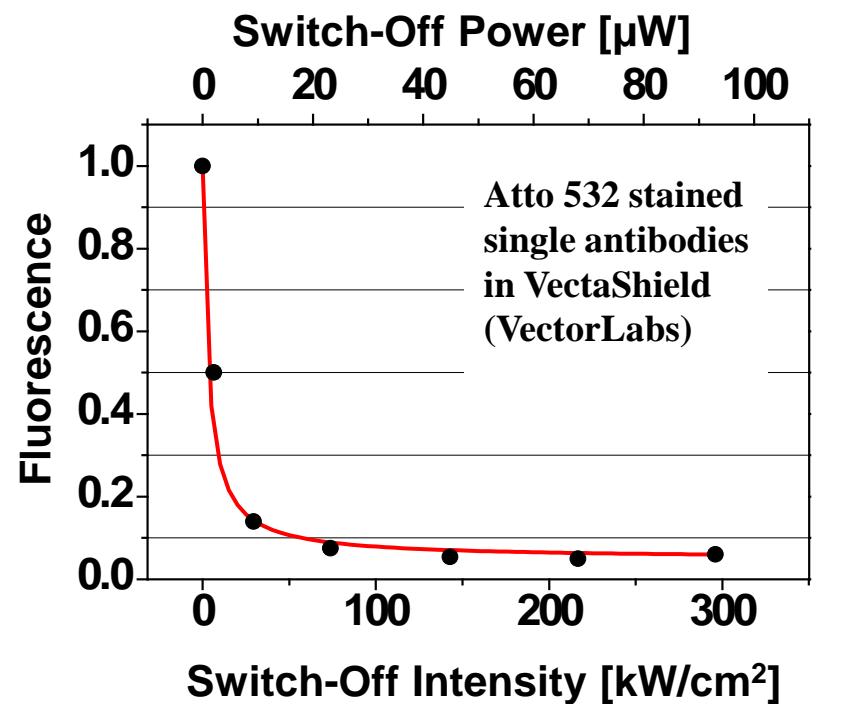
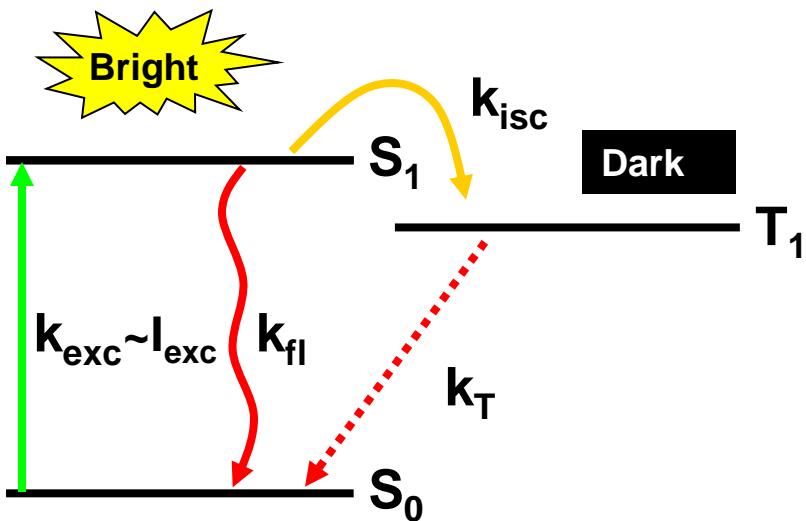


Microtubulin-Peroxisomes, Fölling et al, Nature Methods 2008

Far-Field Nanoscopy

ON/OFF via Triplet/Dark States

GSD (Ground State Depletion)

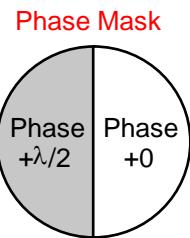


Turn-off fluorescence by pumping
into a long-living dark (triplet) state

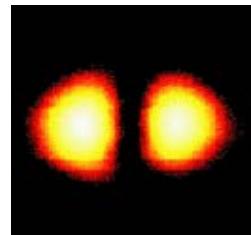
Low CW powers ($\mu\text{W} - \text{kW}/\text{cm}^2$)

GSD-Microscopy

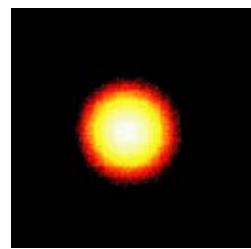
Far-Field Nanoscopy using the triplet state



Switch-off PSF

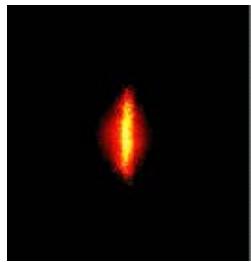


+

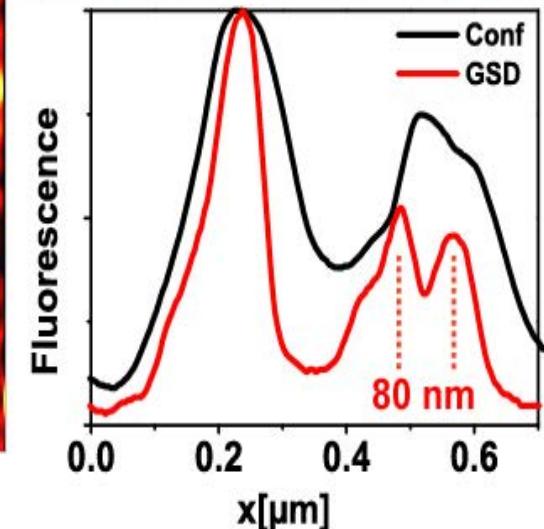
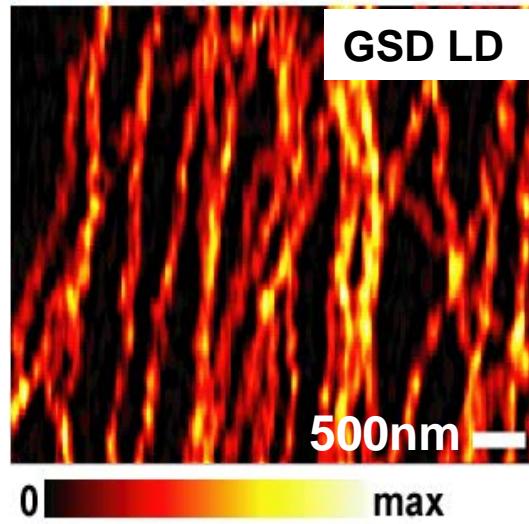
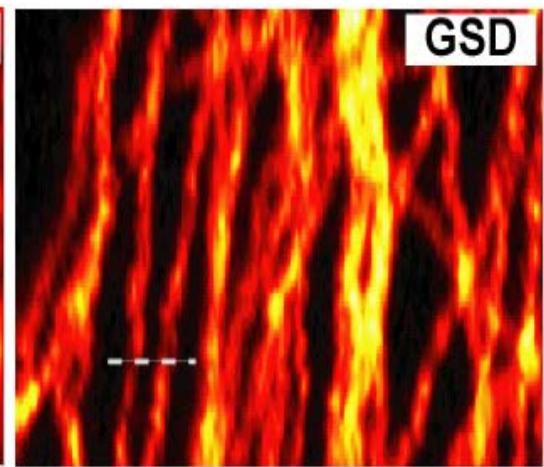
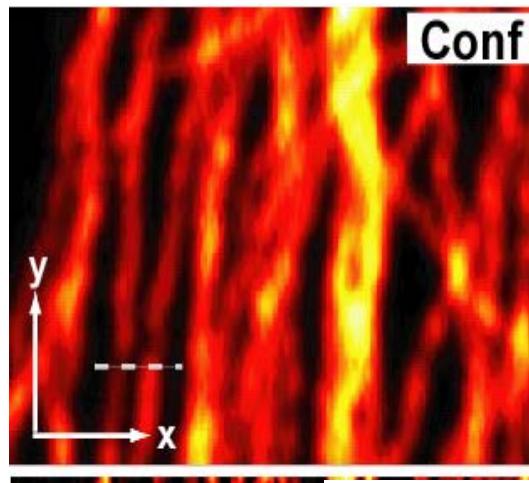


||

Eff. PSF

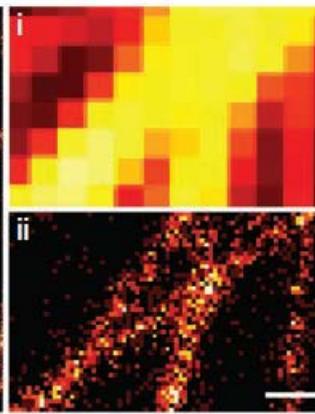
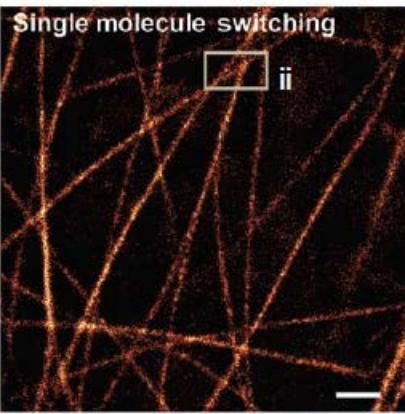
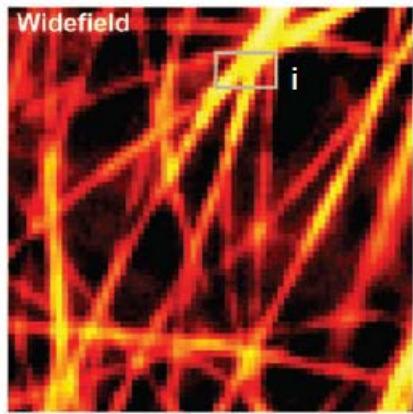


Atto 532 stained microtubuli

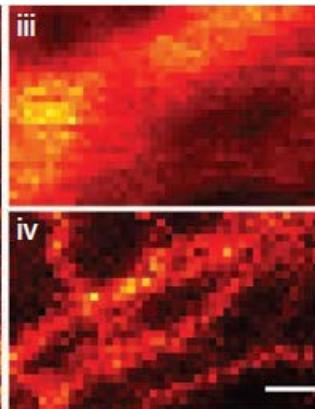
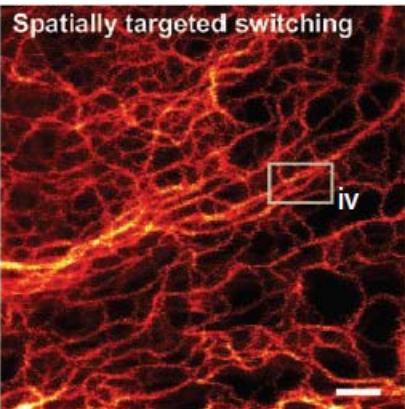
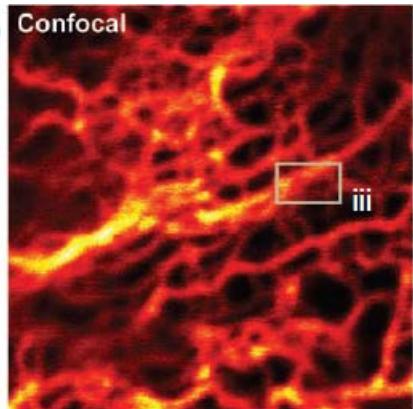


Far-Field Nanoscopy

STED/RESOLFT vs. PALM/STORM/...



PALM/STORM ...



STED/RESOLFT

STED/RESOLFT vs. PALM/...

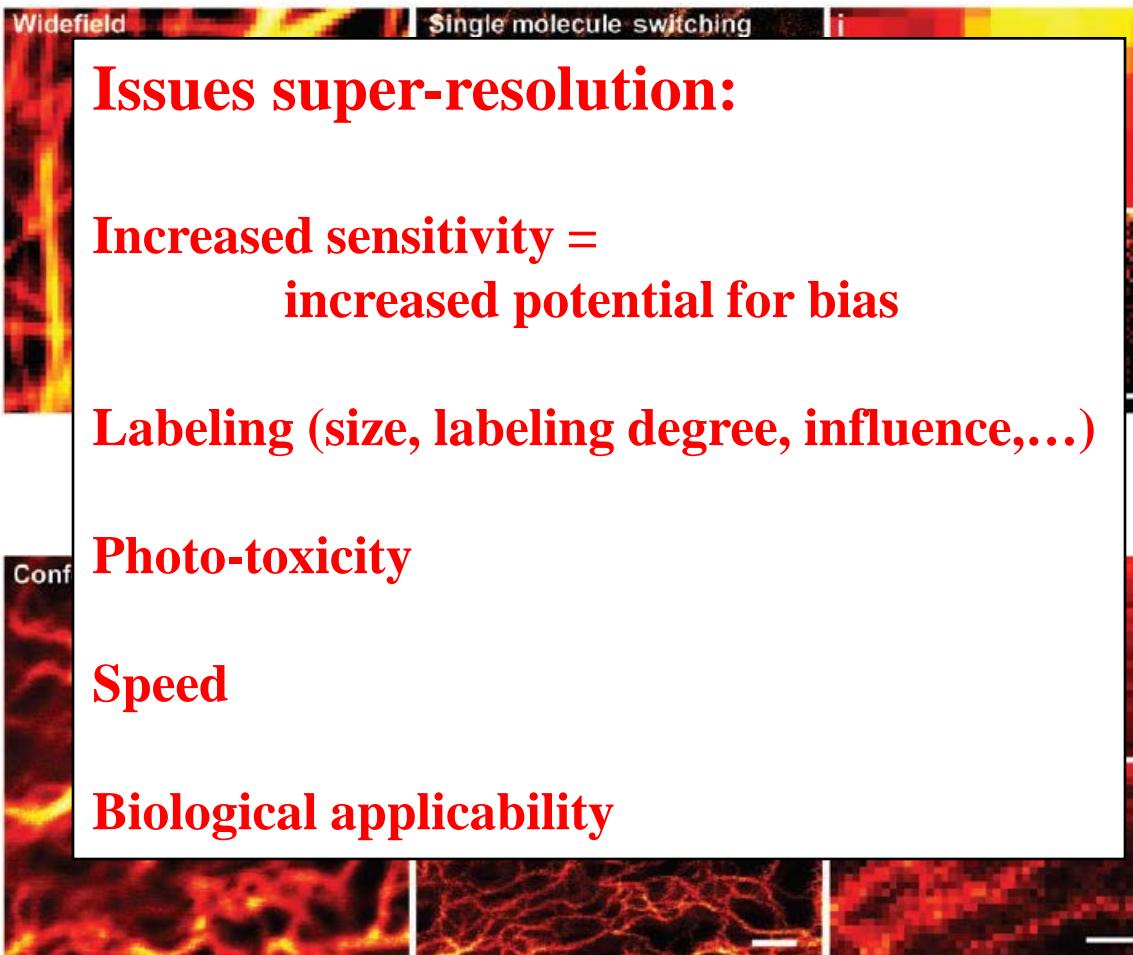
Same principle: ON/OFF

**Similar techniques:
Own advantages/disadvantages**

**Same labels / samples
- New control!**

Far-Field Nanoscopy

STED/RESOLFT vs. PALM/STORM/...



Issues super-resolution:

Increased sensitivity =
increased potential for bias

Labeling (size, labeling degree, influence,...)

Photo-toxicity

Speed

Biological applicability

STED/RESOLFT vs. PALM/...

Same principle: ON/OFF

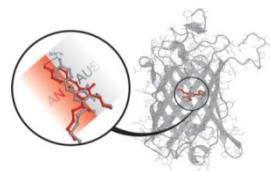
**Similar techniques:
Own advantages/disadvantages**

**Same labels / samples
- New control!**

Far-Field RESOLFT Nanoscopy

Reversibly Photoswitchable Fluorescent Proteins

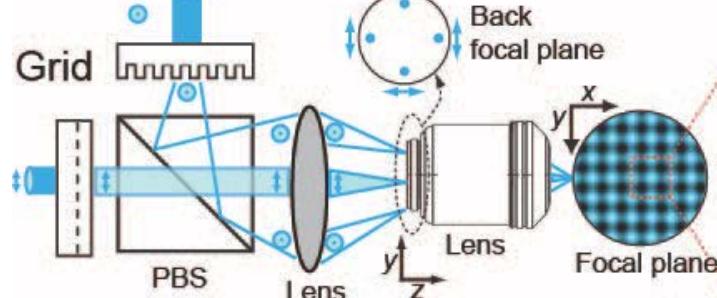
RESOLFT



ON $\xleftrightarrow{\text{light}}$ OFF

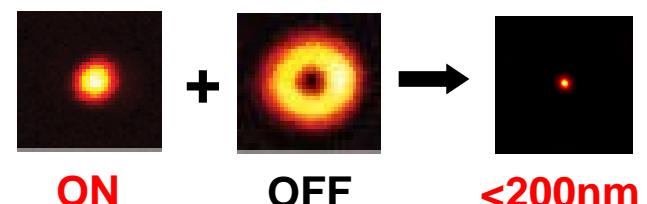
Intensity $\approx 1 \text{ kW/cm}^2$

Parallelization

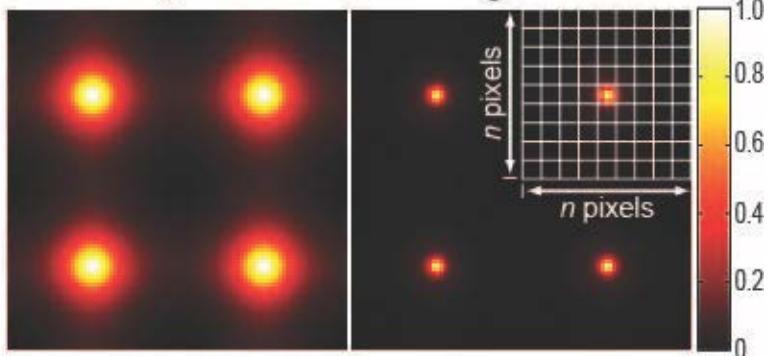


$$I_{\text{off}} = I \cdot \sin^2(x) + I \cdot \sin^2(y)$$

$$I/I_s = 5$$



<200nm



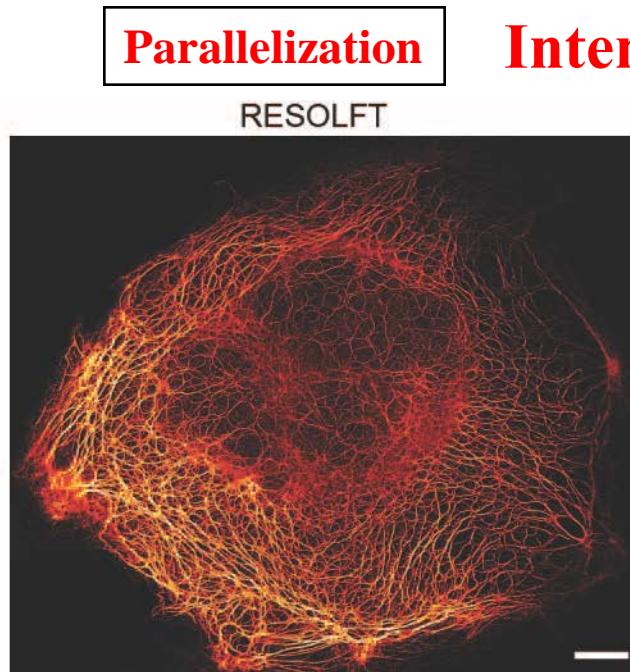
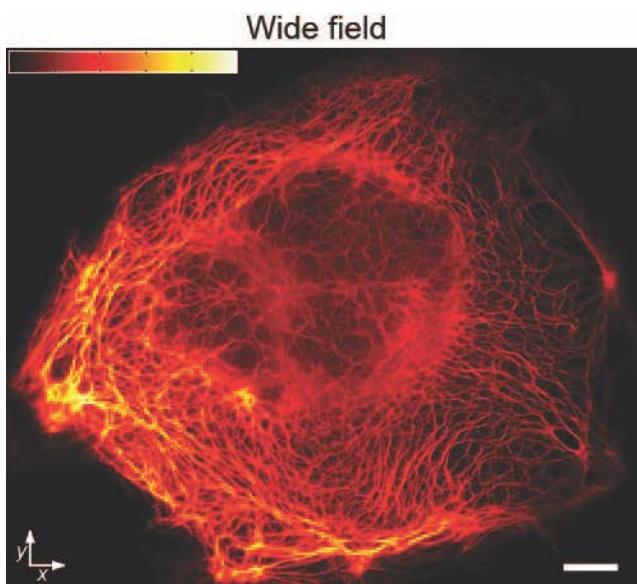
RESOLFT: on-state regions

Thousand doughnuts (CCD detection)

Chmyrov et al, Nature Meth. 2013

Far-Field RESOLFT Nanoscopy

Reversibly Photoswitchable Fluorescent Proteins



Intensity $\approx 1 \text{ kW/cm}^2$

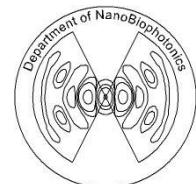
Keratin19-rsEGFP
expressed
in living PtK2 cells

Scale 10 μm

120x100 μm – 1s
Chmyrov et al,
Nature Meth. 2013



Acknowledgement



MPI, Göttingen

Lipid Experiments

Veronika Mueller

Alf Honigmann

Debora Machado Andrade

Christian Ringemann

Rebecca Medda

Birka Lalkens

Giuseppe Viccidomini

Haisen Ta

Andreas Schöngle

Lipid labeling

Dr. V. Belov

S. Polyakova

Stefan Hell

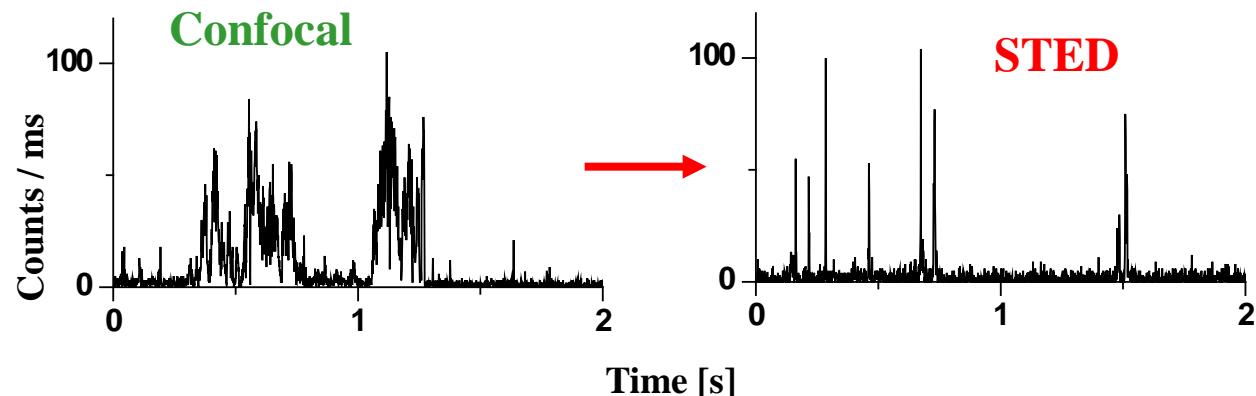
+ whole group

Lipid labeling

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SLB/Phase Sep.

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Jens Ehrig

Petra Schwille
(Dresden)

Membranes

Alf Honigmann

Richard Wagner
(Osnabrück)

Hopping

Aki Kusumi
(Kyoto)

Proteins/Discussions

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Uenal Coskun

Michal Gryzbeck

Daniel Lingwood

Kai Simons

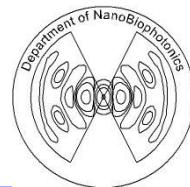
(Dresden)

Fruitful discussion

Herve Rigneault
(Marseille)



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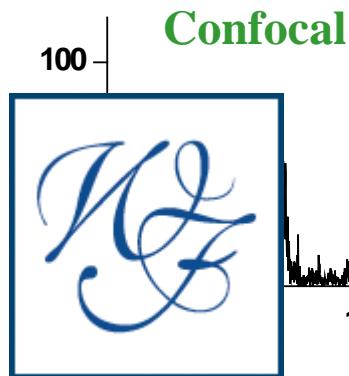
S. Polyakova

Lipid labeling

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**Stefan Hell
+ whole group**

Team – HIU/WIMM:

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Mathias Clausen (Biophysics - membrane)

Silvia Galiani (Physics – nanoscope setup/organelles)

Marco Fritzsché (Physics - cytoskeleton)

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Doug Higgs

Simon Davis

David Jackson

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Micron

Ilan Davis, Lothar Schermelleh, ...

Martin Booth, Achillefs Kapanidis,

Philipp Kukura...